

## Supporting Information

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### Cytotoxic and Antibacterial Activities of Constituents from *Calophyllum ferrugineum* Ridley

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## Experimental Details

### *Cytotoxic Activity:*

The cytotoxic activity was evaluated by MTT colorimetric assay [1,2]. The sample stock solution (100 µg/mL) was dissolved in 1% (v/v) DMSO in phosphate buffered saline (PBS). The samples were further diluted with DMEM to afford concentration ranging from 100 – 3.13 µg/mL obtained from twofold dilution. The cells were cultured in Dulbecco's modified Eagle's Medium (DMEM) media supplemented with 10% fetal bovine serum and 2% penicillin-streptomycin. In brief, 90 µL of cell suspension in DMEM were seeded in 96-well microplate and was counted directly by using trypan blue dye. The cells were treated with samples after reaching confluence ( $2 \times 10^5$  cell/mL) and were pre-incubated at 37°C in humidified atmosphere with 5% CO<sub>2</sub> for 24 hours. 20 µL of MTT (5 mg/mL in PBS) was added to all well in dark condition and pre-incubated for another 4 hours. 100 µL of DMSO was added to all well to solubilize the water-insoluble purple formazan crystal formed and pre-incubated in dark condition at room temperature. The absorbance was read after 1 hour at 570 nm and 630 nm as the reference wavelength. Untreated cells served as control group and considered as 100% of viable cells. Results were expressed as percentage of cell viability of samples relative to the untreated control cell following the formula;

$$\% \text{Inhibition Concentration (\% IC)} = [(A_{\text{sample}} - A_{\text{MTT blank}}) / (A_{\text{control}} - A_{\text{MTT blank}})] \times 100\%$$

where  $A_{\text{sample}}$  is the absorbance of cells treated with samples,  $A_{\text{MTT blank}}$  is the absorbance of MTT reagent with DMSO only and  $A_{\text{control}}$  is the absorbance of untreated control cells.

### *Antibacterial Activity:*

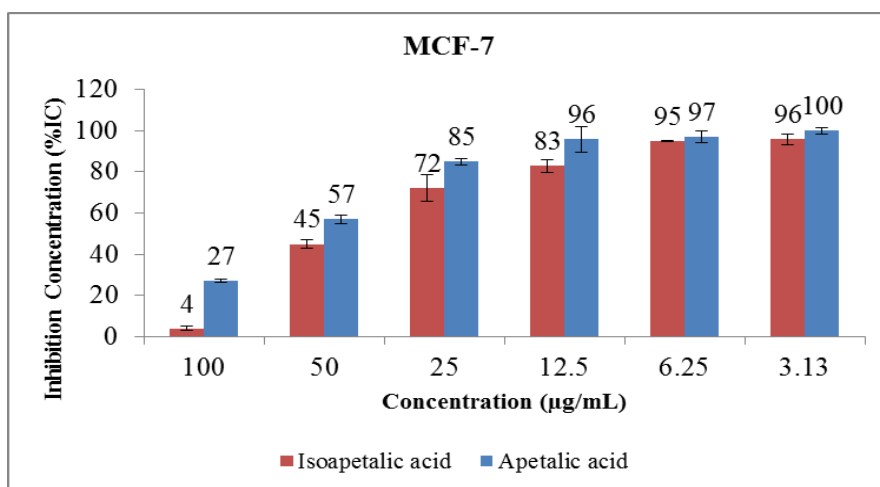
The antibacterial activity of all compounds was tested quantitatively by evaluating their minimum inhibition concentration (MIC). The MIC was carried out by micro-broth dilution [3–7]. The sample stock solution (1000 µg/mL) was prepared in 5% DMSO in nutrient broth (NB) supplemented with 0.02% (v/v) Tween 80. Further twofold dilution with NB was performed to afford concentration of samples from 100 – 7.81 µg/mL. 50 µL of bacteria inocula ( $10^6$  CFU/mL) was dispensed in the 96-well microplate followed by 50 µL of the sample solution. The microplates were pre-incubated for 24 hour at 37°C for *S. aureus*, *E. coli* and *P. aeruginosa* and 30 °C for *B. subtilis*. 25 µL of 2-(4-Iodophenyl)-3-(4-nitrophenyl)-5-phenyl-2H-tetrazolium (INT) (0.2 mg/mL in distilled water) solution was added to all wells and were further pre-incubated for at least 30 minutes. Bacteria growth in the wells was indicated by formation of reddish-pink colour while clear well indicates inhibition of bacteria growth by the sample. Streptomycin sulphate was employed as positive control in this assay.

### *Statistical Analysis of Data:*

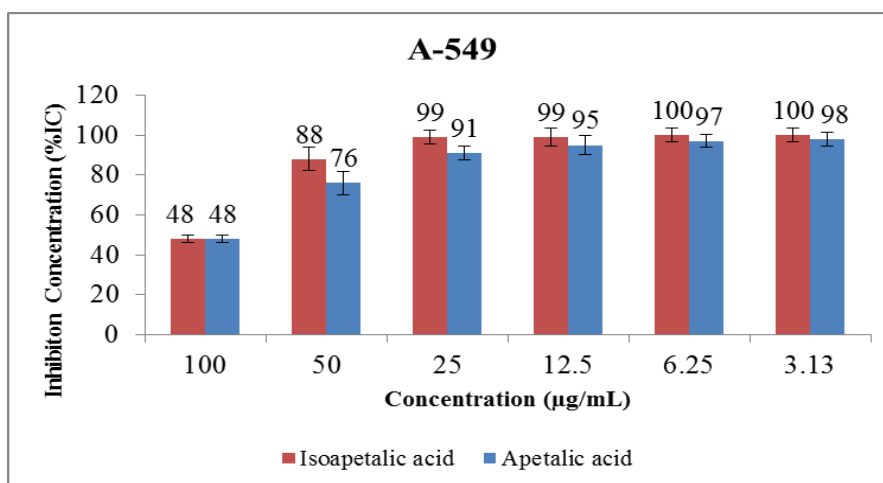
Three replicates of each sample were used for statistical analysis with values reported as mean ± SD. Standard curves were generated and calculation of the 50% inhibitory concentration (IC<sub>50</sub>) values was

performed using GraphPad Prism for Windows (version 5.02) software. The Student's *t*-test was carried out using SPSS (version 22) software to study the comparison between treatment of samples and untreated control. A value of  $p < 0.05$  was considered significantly different.

**Bar Graph on Cytotoxic Activity of Isoapetalic acid (1) and Apetalic acid (2) against A-549 and MCF-7 cell lines at six different concentrations**

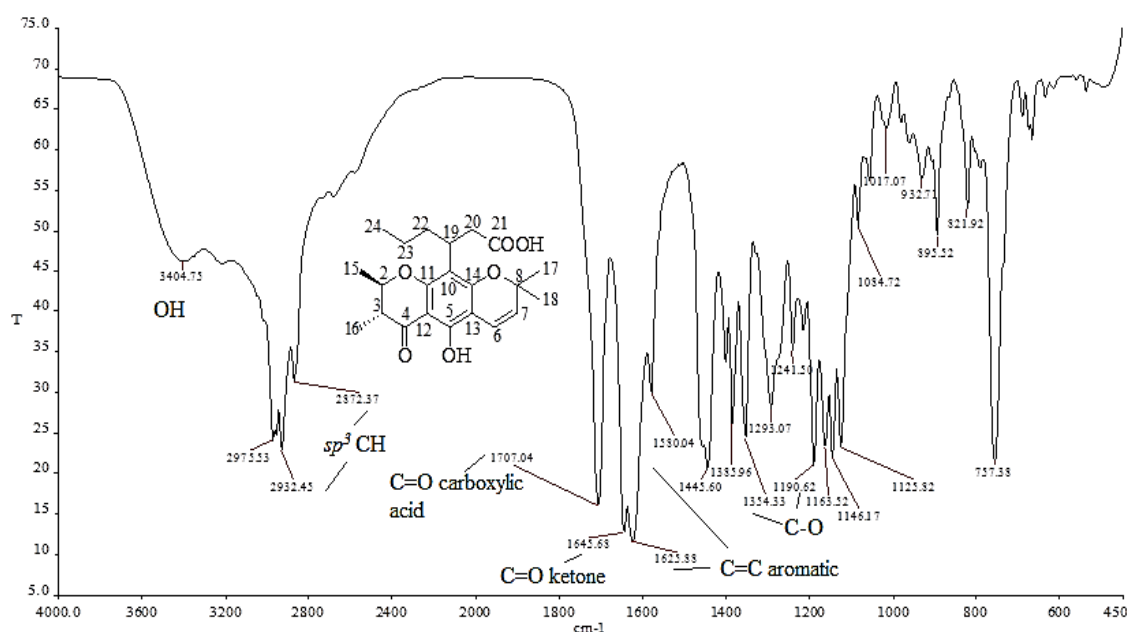


Percentage of Inhibition Concentration (%IC) of Isoapetalic acid (1) and Apetalic acid (2) against MCF-7

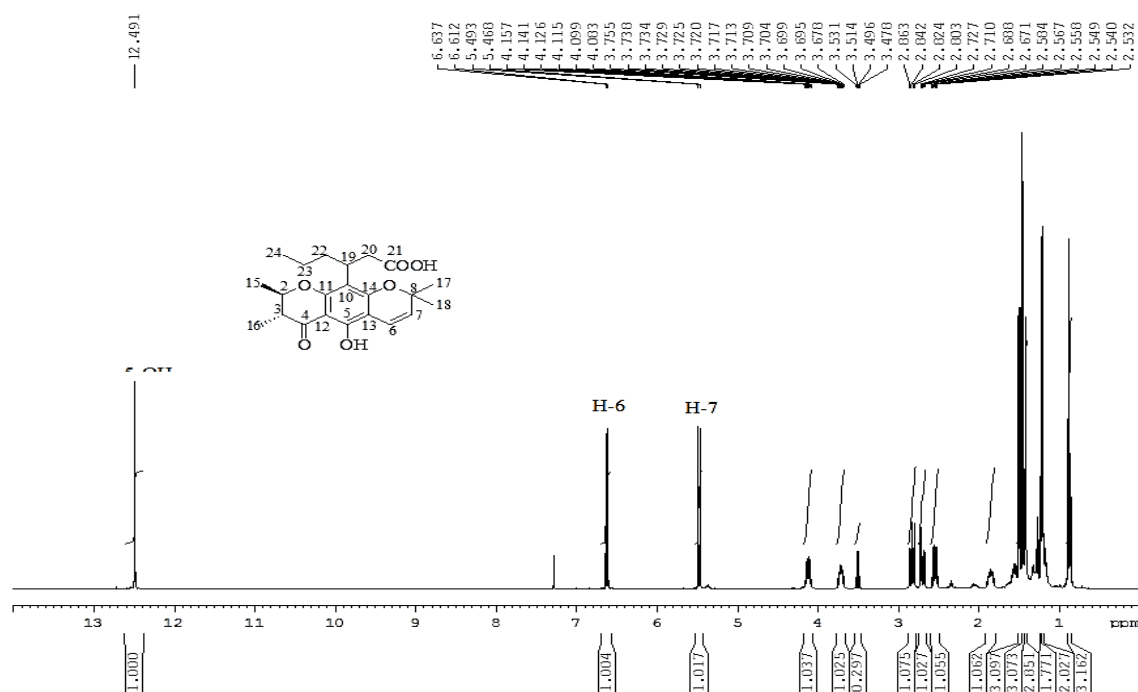


Percentage of Inhibition Concentration (%IC) of Isoapetalic acid (1) and Apetalic acid (2) against A-549

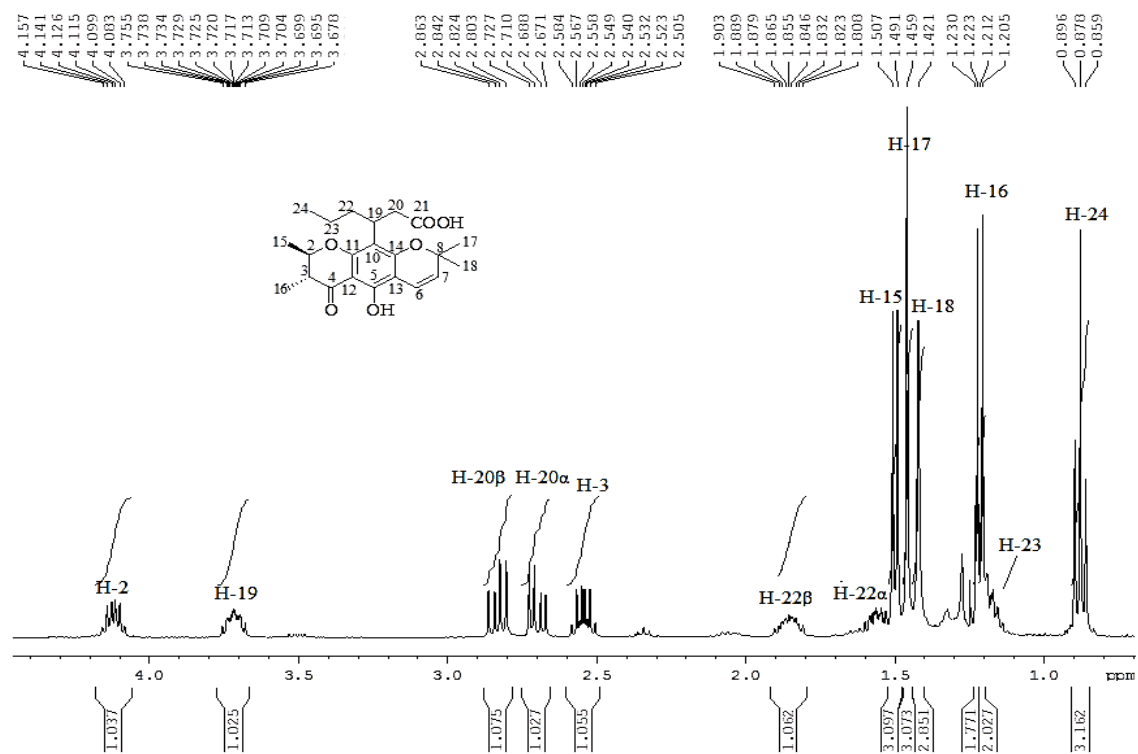
Isoapetalic acid (**1**): Pale yellow gum;  $R_f$  0.38 (*n*-Hex:Et<sub>2</sub>O, 1:1);  $[\alpha]_D^{25}$  -196.7° (*c* 0.033, CHCl<sub>3</sub>); IR (NaCl disc, CHCl<sub>3</sub>)  $\nu_{\max}$  cm<sup>-1</sup>: 3405 (OH), 2975 and 2932 (*sp*<sup>3</sup>CH), 1707 (C=O acid), 1646 (chelate C=O ketone), 1626 and 1580 (C=C aromatic), 1354 and 1190 (C-O); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  0.88 (3H, t, *J* = 7.2 Hz, H-24), 1.20 (2H, m, H-23), 1.22 (3H, d, *J* = 7.2 Hz, H-16), 1.42 (3H, s, H-18), 1.46 (3H, s, H-17), 1.50 (3H, d, *J* = 6.4 Hz, H-15), 1.58 (1H, m, H-22 $\alpha$ ), 1.86 (1H, m, H-22 $\beta$ ), 2.55 (1H, dq, *J* = 10.8 and 7.2 Hz, H-3), 2.68 (1H, dd, *J* = 15.6 and 6.8 Hz, H-20 $\alpha$ ), 2.83 (1H, dd, *J* = 15.2 and 8.4 Hz, H-20 $\beta$ ), 3.71 (1H, m, H-19), 4.12 (1H, dq, *J* = 10.8 and 6.4 Hz, H-2), 5.48 (1H, d, *J* = 10.0 Hz, H-7), 6.62 (1H, d, *J* = 10.0 Hz, H-6) and 12.49 (1H, s, 5-OH); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta$  10.5 (C-16), 14.1 (C-24), 19.5 (C-15), 20.9 (C-23), 28.2 (C-17), 28.4 (C-18), 30.4 (C-19), 35.5 (C-22), 38.6 (C-20), 45.8 (C-3), 78.1 (C-8), 78.9 (C-2), 101.9 (C-12), 102.6 (C-13), 109.0 (C-10), 115.7 (C-6), 125.6 (C-7), 157.0 (C-5), 159.9 (C-11 and C-14), 179.0 (C-21) and 199.4 (C-4); EIMS (% rel int): *m/z* 388 (12), [M]<sup>+</sup> (C<sub>22</sub>H<sub>28</sub>O<sub>6</sub>), 373 (100), 329 (5).



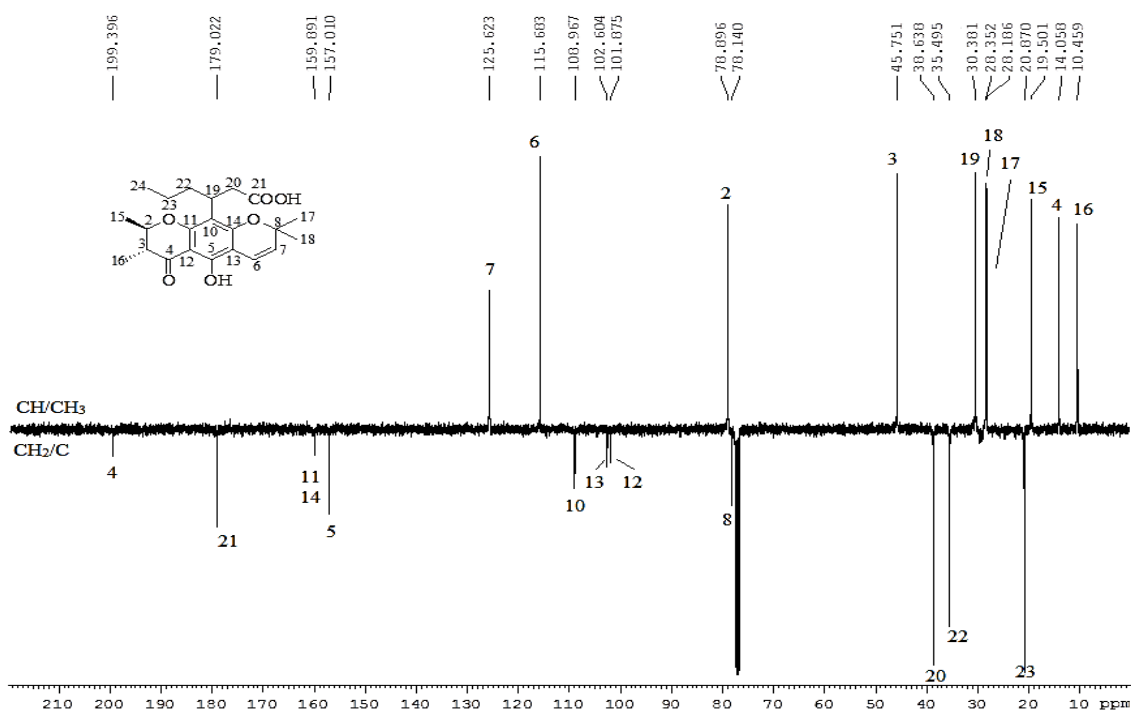
S1: IR spectrum of Isoapetalic acid (**1**)



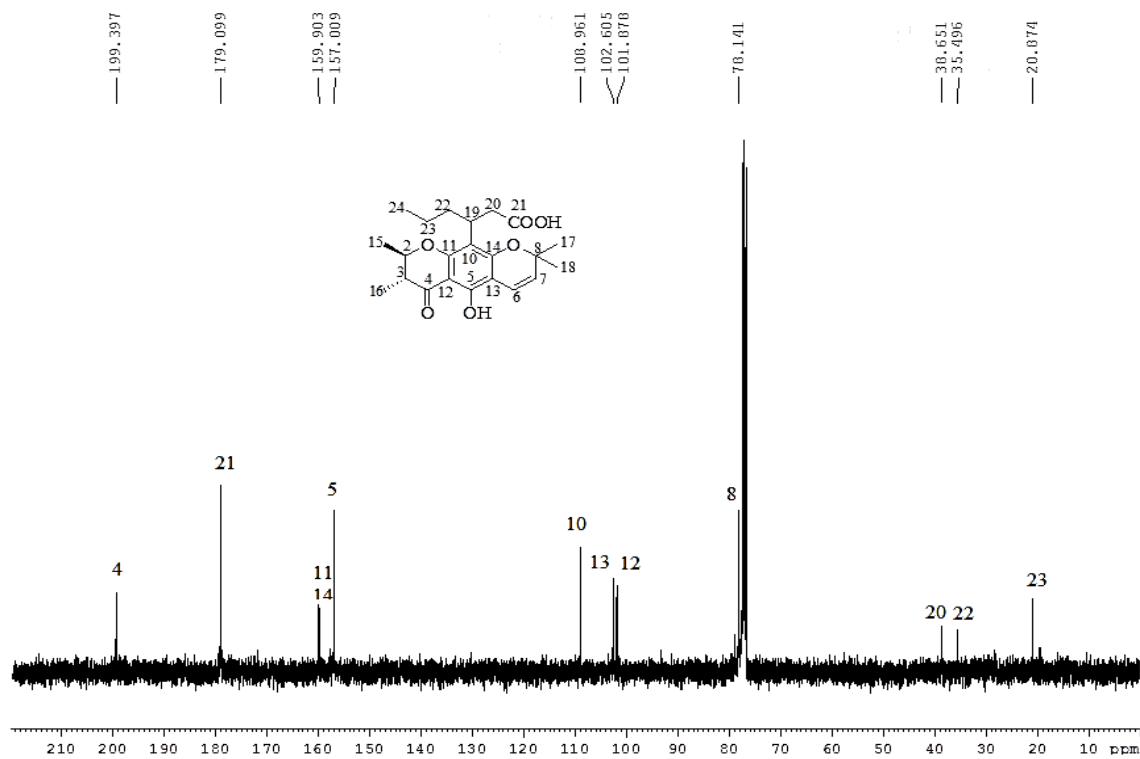
**S2:** <sup>1</sup>H NMR spectrum of Isoapetalic acid (1)



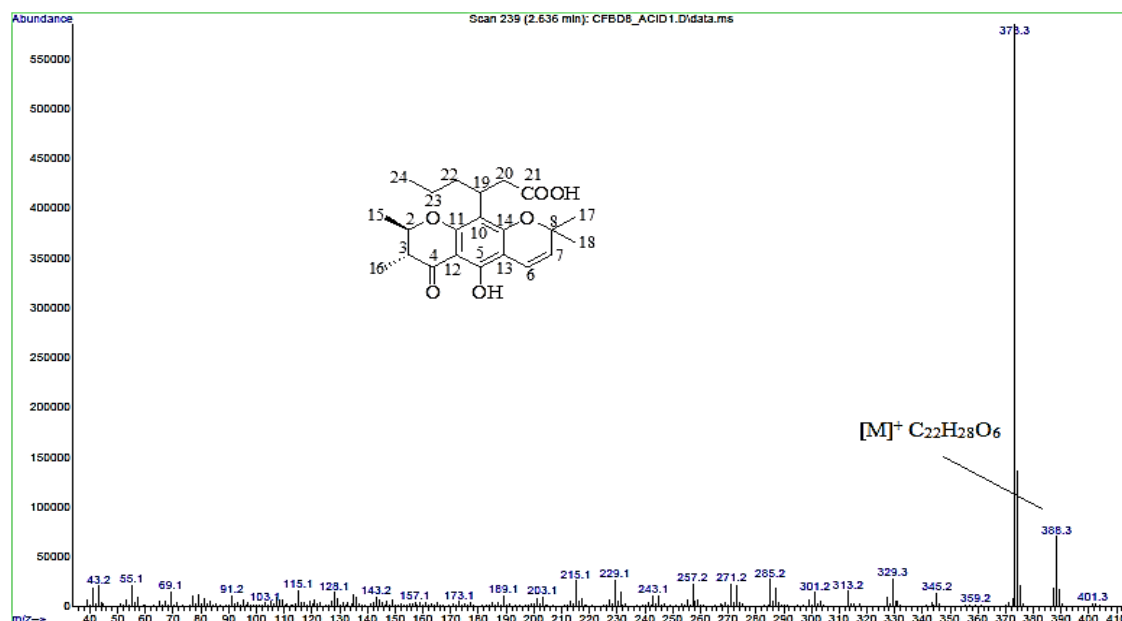
**S3:** <sup>1</sup>H NMR spectrum of Isoapetalic acid (1) (Expansion)



S4: DEPTQ spectrum of Isoapetalic acid (1)



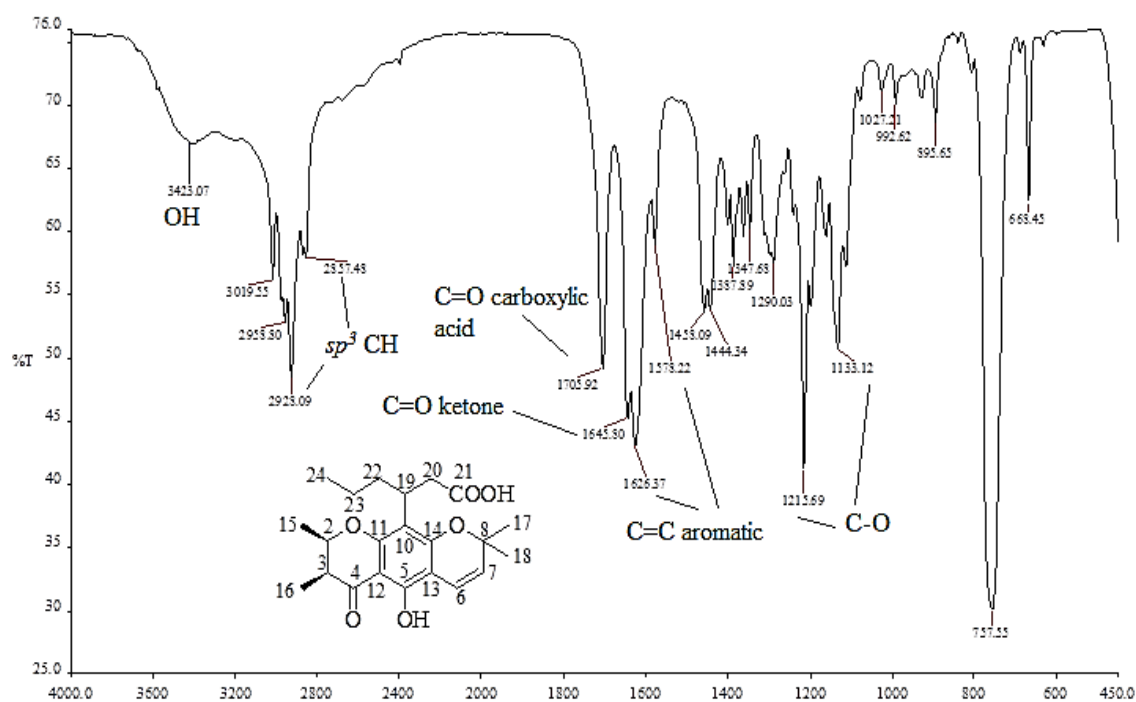
S5: DEPTQ\_Q spectrum of Isoapetalic acid (1)



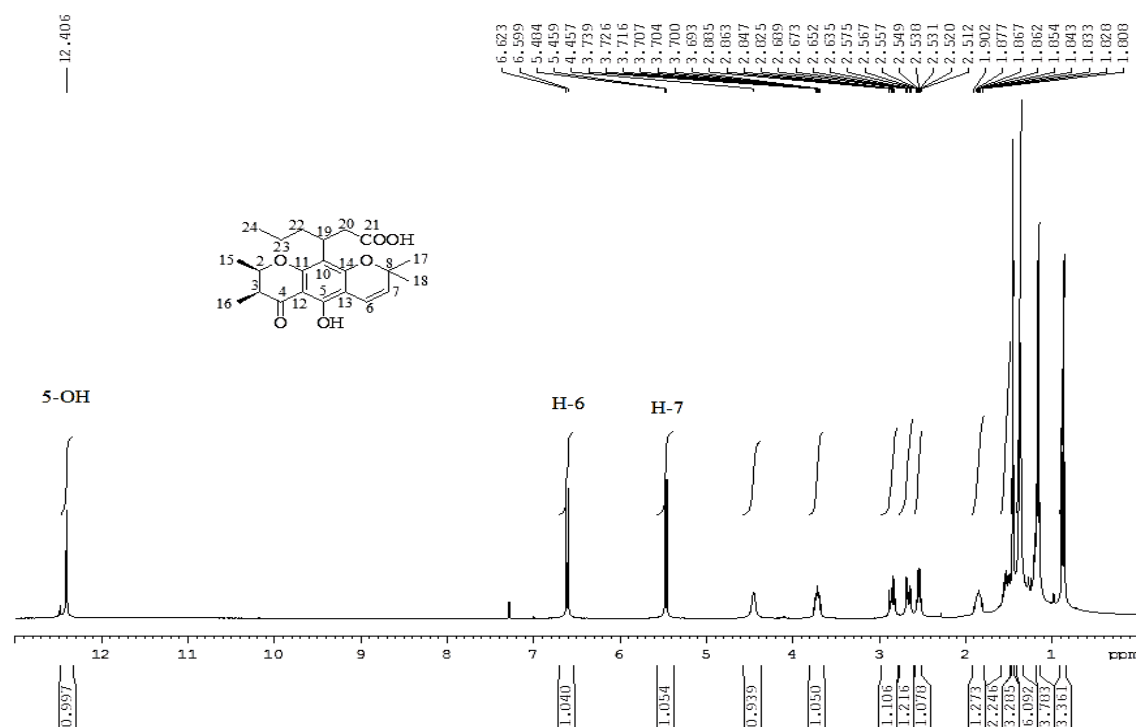
**S6:** EIMS spectrum of Isoapetalic acid (**1**)

Apetalic acid (**2**): Yellow gum;  $R_f$  0.25 (*n*-Hex:Et<sub>2</sub>O, 1:1);  $[\alpha]_D^{25}$  -73.4° (*c* 0.033, CHCl<sub>3</sub>); IR (NaCl disc, CHCl<sub>3</sub>)  $\nu_{\max}$  cm<sup>-1</sup>: 3423 (OH), 2928 and 2857 (*sp*<sup>3</sup> CH), 1705 (C=O acid), 1646 (cholate C=O ketone), 1626 and 1578 (C=C aromatic), 1215 and 1133 (C-O); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  0.87 (3H, t, *J* = 7.2 Hz, H-24), 1.16 (3H, d, *J* = 7.2 Hz, H-16), 1.20 (2H, m, H-23), 1.37 (3H, d, *J* = 6.4 Hz, H-15), 1.38 (3H, s, H-18), 1.46 (3H, s, H-17), 1.58 (1H, m, H-22 $\alpha$ ), 1.86 (1H, m, H-22 $\beta$ ), 2.55 (1H, qd, *J* = 7.2 and 3.2 Hz, H-3), 2.66 (1H, dd, *J* = 15.6 and 6.8 Hz, H-20 $\alpha$ ), 2.85 (1H, dd, *J* = 15.2 and 8.4 Hz, H-20 $\beta$ ), 3.71 (1H, m, H-19), 4.46 (1H, br s, H-2), 5.47 (1H, d, *J* = 10.0 Hz, H-7), 6.61 (1H, d, *J* = 10.0 Hz, H-6) and 12.41 (1H, s, 5-OH); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>):  $\delta$  9.3 (C-16), 14.0 (C-24), 16.3 (C-15), 20.8 (C-23), 28.1 (C-17), 28.4 (C-18), 30.5 (C-19), 35.5 (C-22), 38.6 (C-20), 44.2 (C-3), 76.1 (C-2), 78.2 (C-8), 101.2 (C-12), 102.6 (C-13), 108.7 (C-10), 115.6 (C-6), 125.7 (C-7), 157.3 (C-5), 159.9 (C-11 and C-14), 179.4 (C-21) and 201.3 (C-4); EIMS (% rel int): *m/z* 388 (12), [M]<sup>+</sup> (C<sub>22</sub>H<sub>28</sub>O<sub>6</sub>), 373 (100), 329 (5).

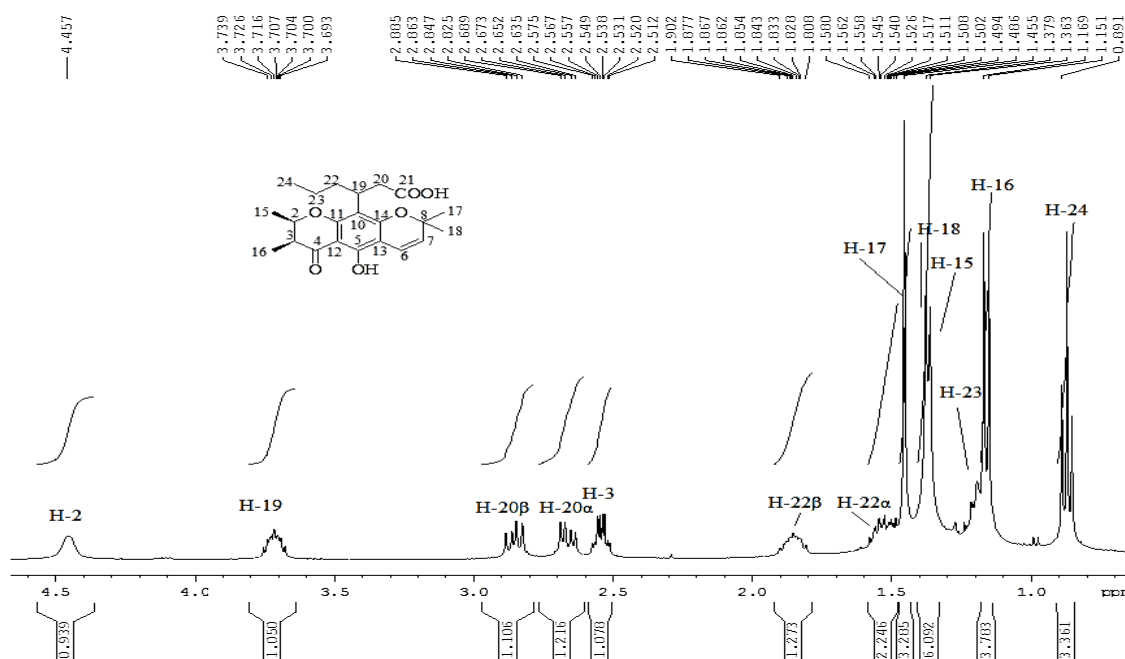




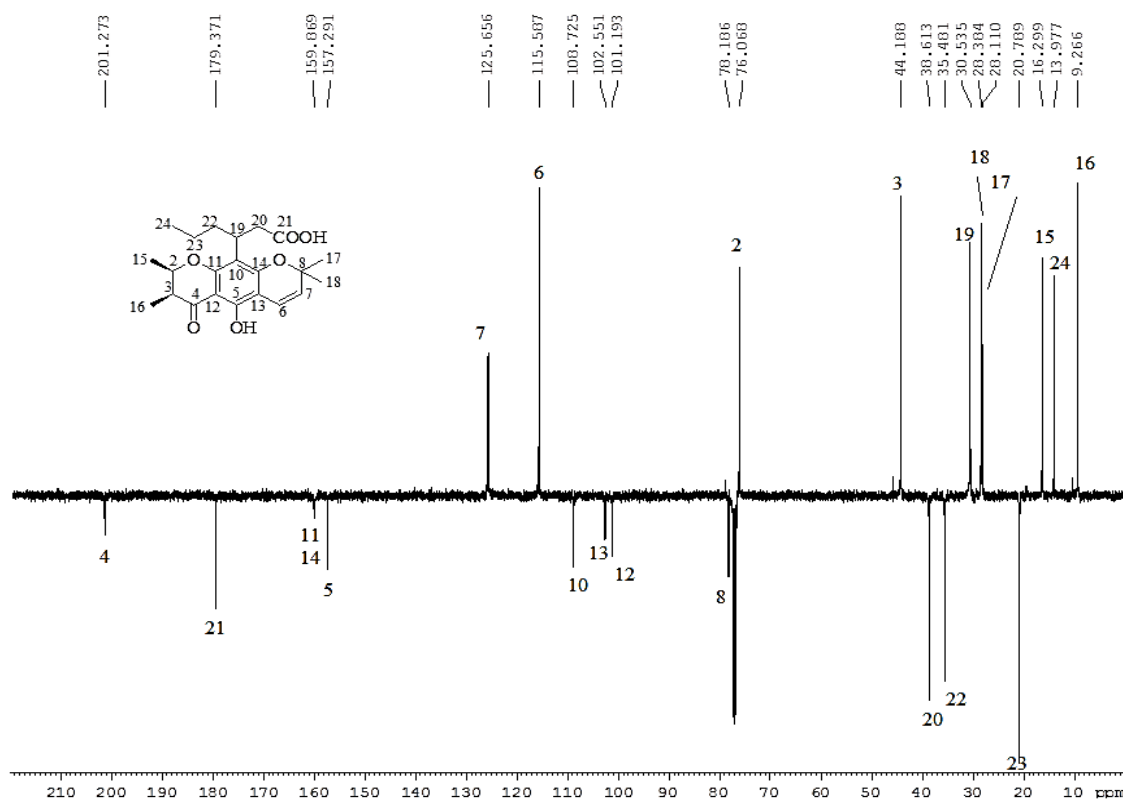
S7: IR spectrum of Apetalic acid (2)



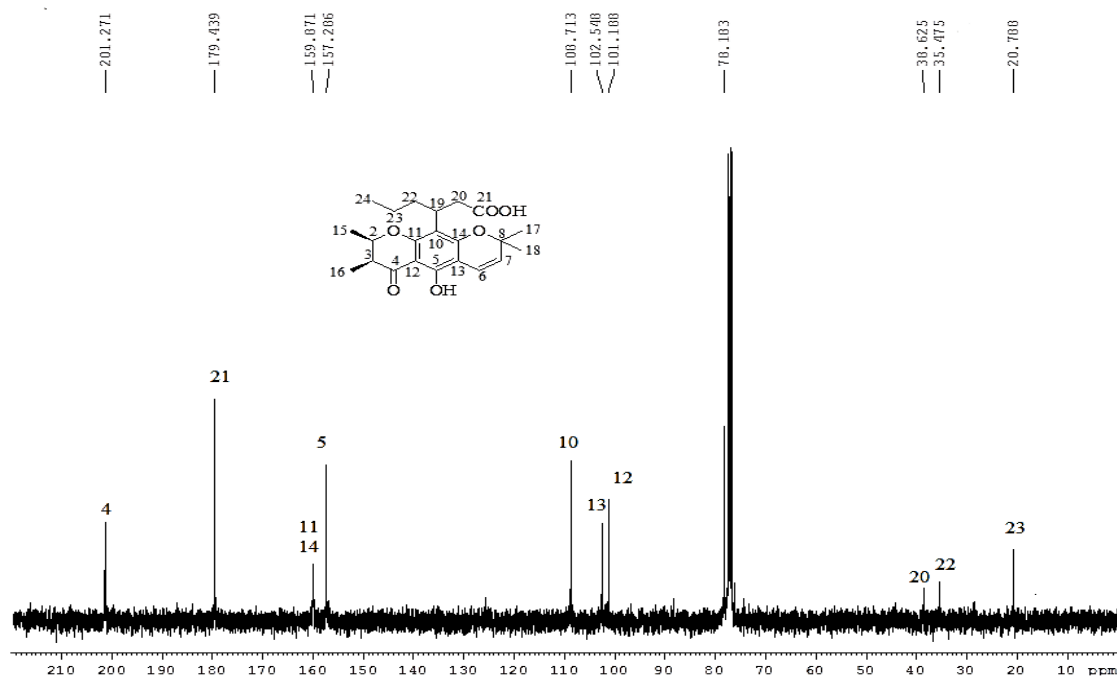
S8:  $^1\text{H}$  NMR spectrum of Apetalic acid (2)



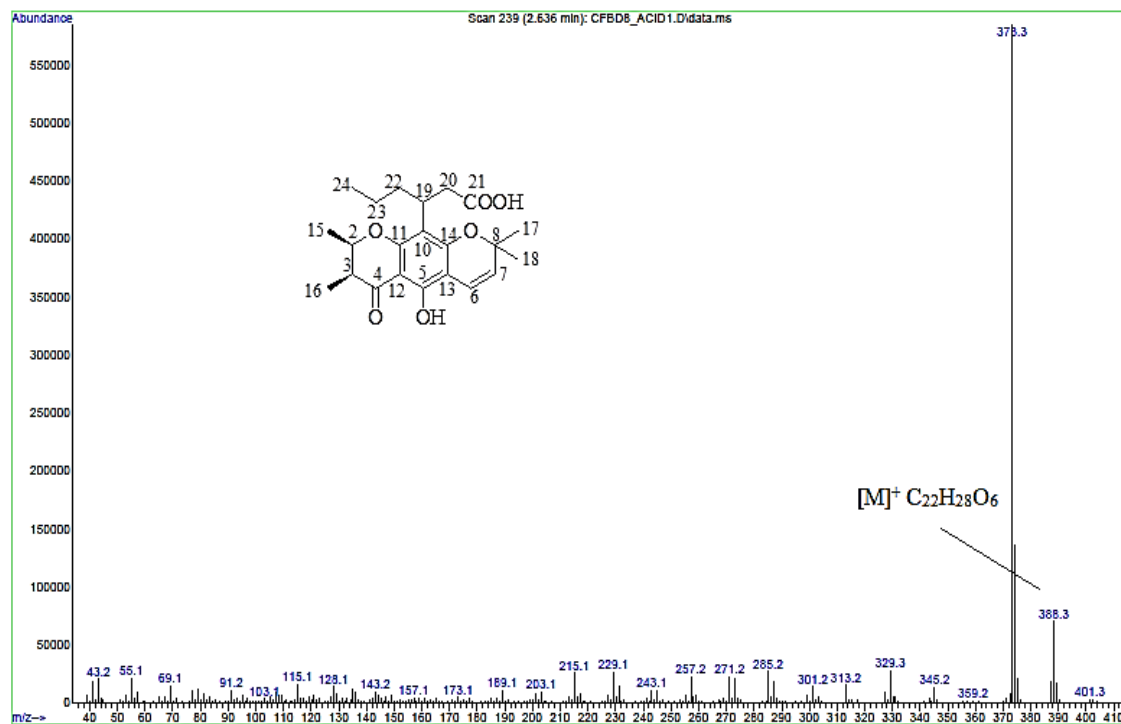
**S9:  $^1\text{H}$  NMR spectrum of Apetalic acid (2) (Expansion)**



**S10: DEPTQ spectrum of Apetalic acid (2)**

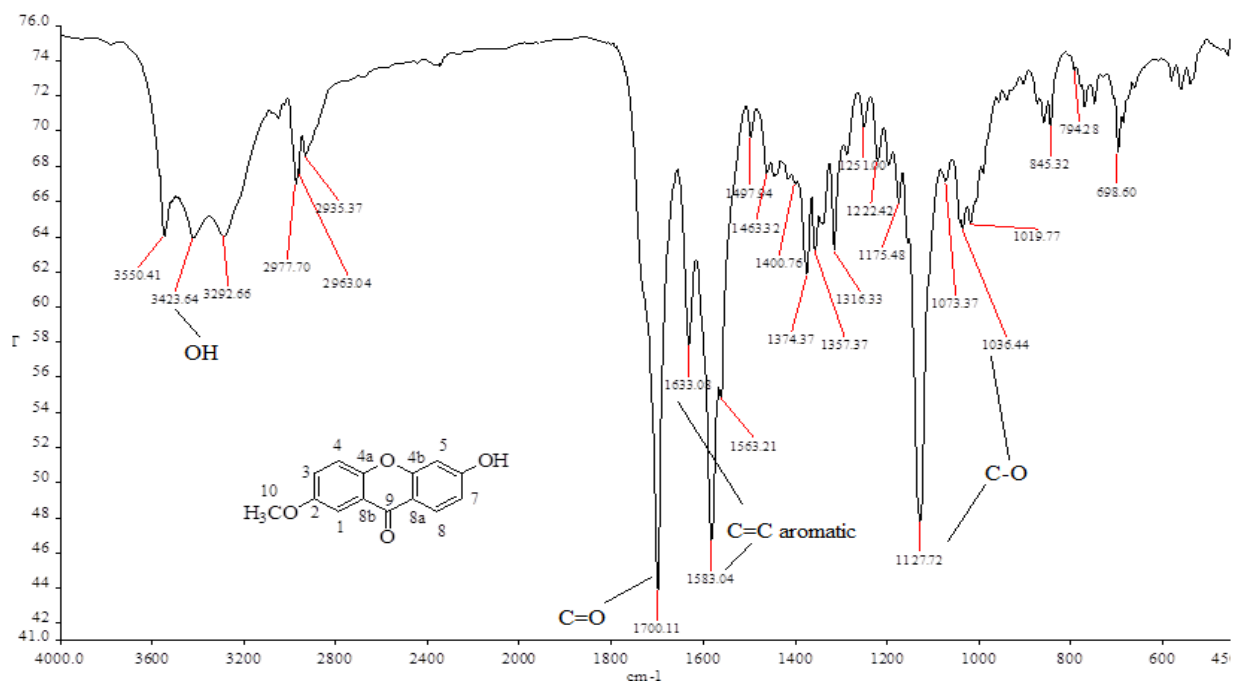


**S11:** DEPTQ-Q spectrum of Apetalic acid (2)

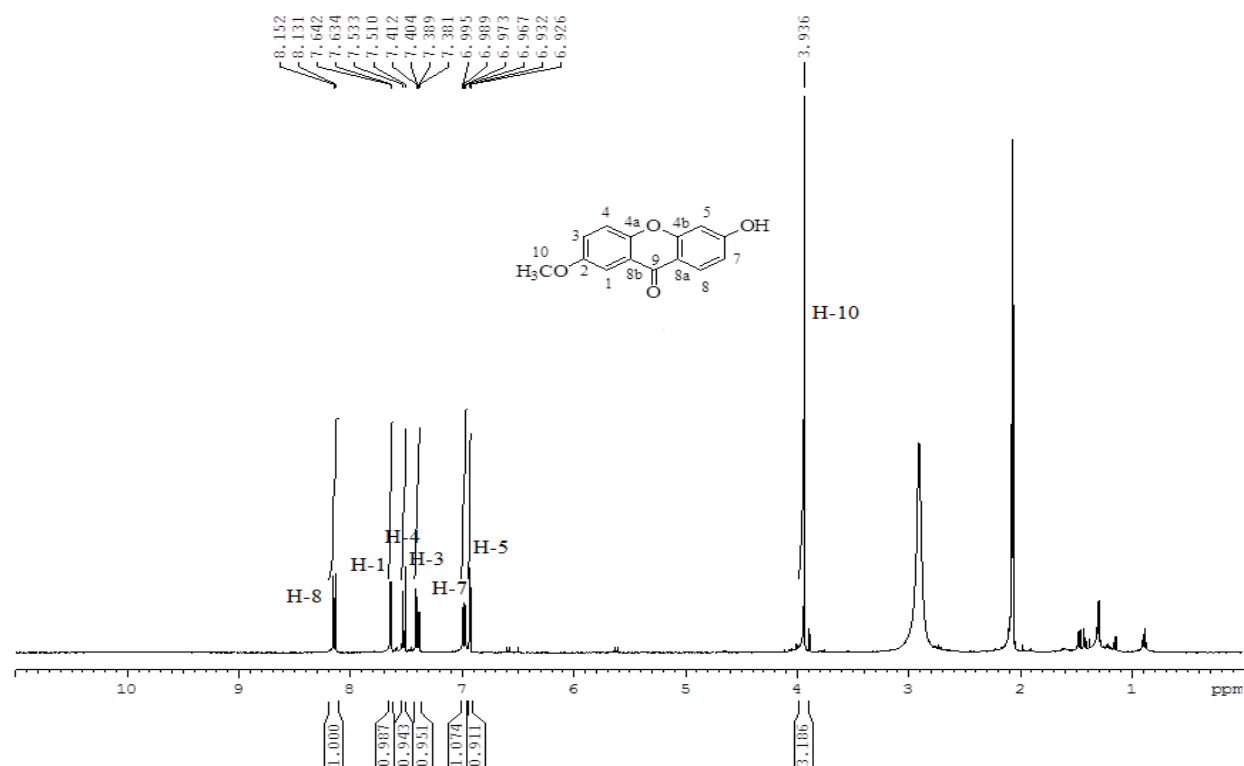


**S12:** EIMS spectrum of Apetalic acid (2)

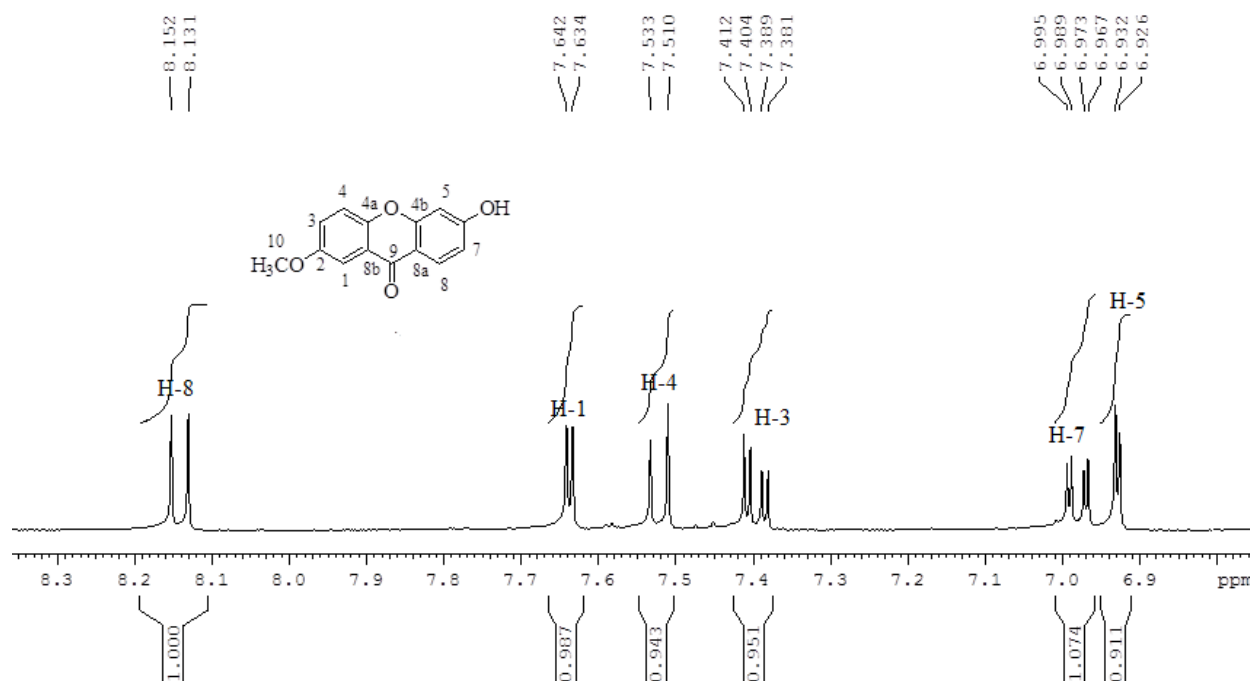
6-Hydroxy-2-methoxyxanthone (**3**): Pale yellow solid;  $R_f$  0.45 (*n*-Hex:EtOAc, 1:1); m.p 263 – 265°C; IR (KBr pellet)  $\nu_{\max}$   $\text{cm}^{-1}$ : 3423 (OH), 1700 (C=O ketone), 1633 and 1583 (C=C aromatic), 1127 and 1036 (C-O);  $^1\text{H}$  NMR (400 MHz,  $\text{CD}_3\text{COCD}_3$ ):  $\delta$  3.94 (3H, s, 10-OCH<sub>3</sub>), 6.93 (1H, d,  $J$  = 2.4 Hz, H-5), 6.98 (1H, dd,  $J$  = 8.8 and 2.4 Hz, H-7), 7.40 (1H, dd,  $J$  = 9.2 and 3.2 Hz, H-3), 7.53 (1H, d,  $J$  = 9.2 Hz, H-4), 7.64 (1H, d,  $J$  = 3.2 Hz, H-1), 8.14 (1H, d,  $J$  = 8.8 Hz, H-8) and 9.93 (1H, s, 6-OH);  $^{13}\text{C}$  NMR (100 MHz,  $\text{CD}_3\text{COCD}_3$ ):  $\delta$  55.3 (C-10), 102.1 (C-5), 106.1 (C-1), 113.8 (C-7), 114.4 (C-8a), 119.2 (C-4), 122.2 (C-8b), 123.5 (C-3), 128.1 (C-8), 150.7 (C-4a), 156.1 (C-2), 158.0 (C-4b), 163.7 (C-6) and 174.9 (C-9); EIMS (% rel int):  $m/z$  242 (100),  $[\text{M}]^+$  ( $\text{C}_{14}\text{H}_{10}\text{O}_4$ ), 227 (24), 212 (31).



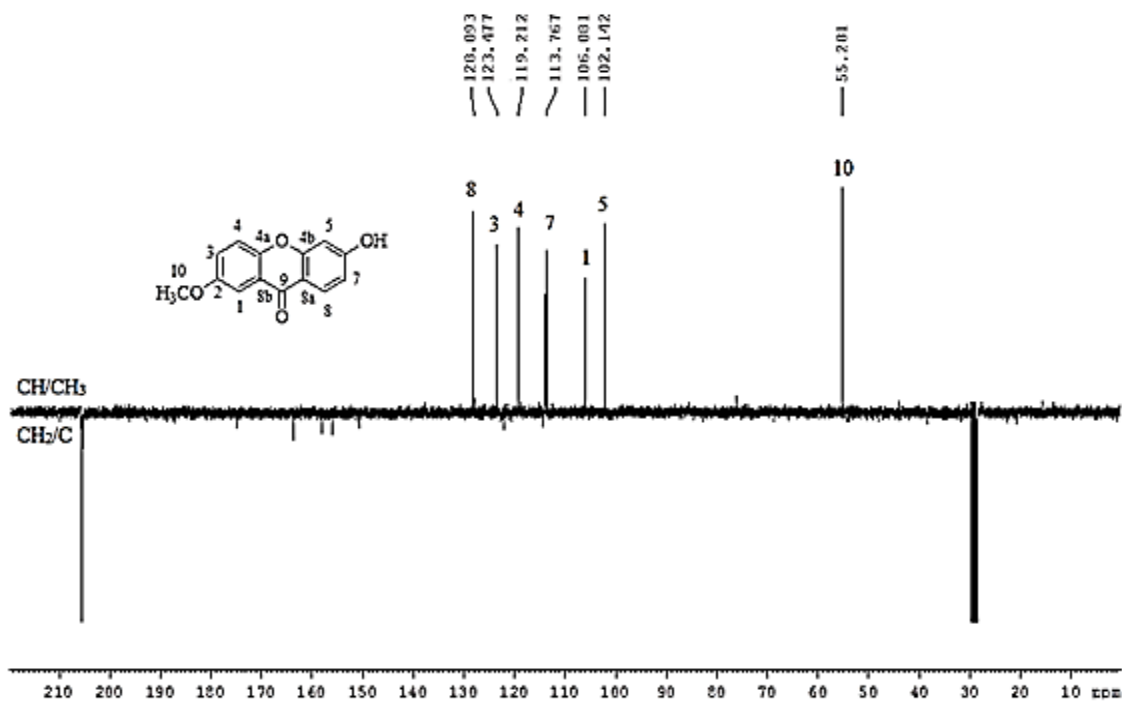
**S13:** IR spectrum of 6-Hydroxy-2-methoxyxanthone (**3**)



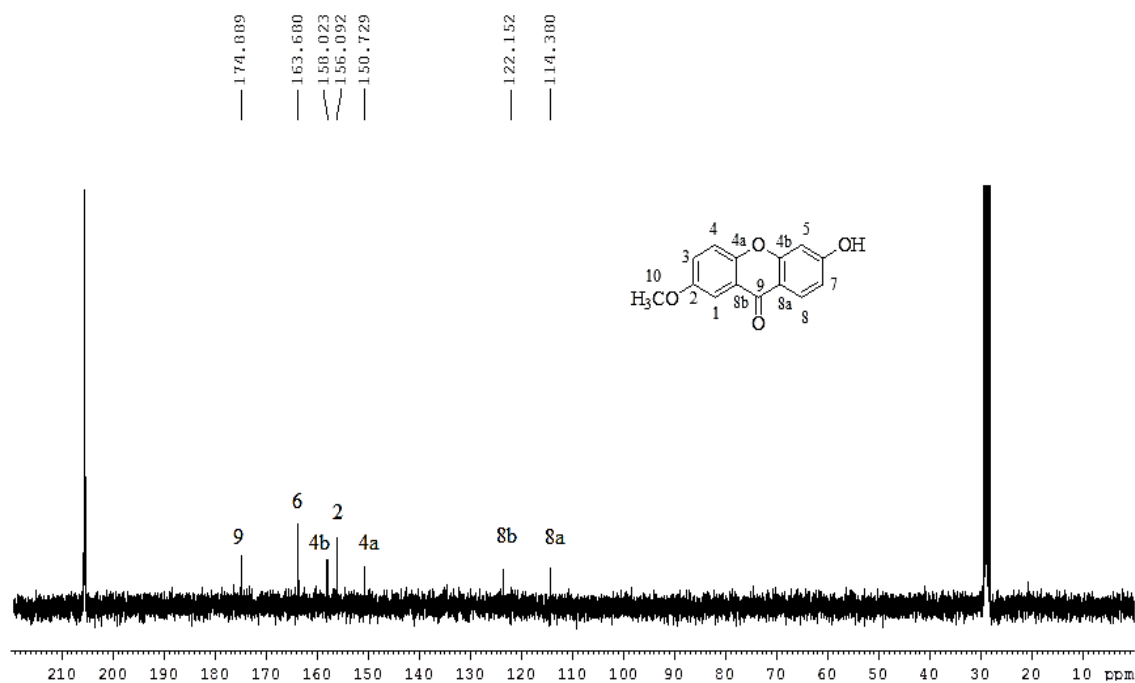
**S14:**  $^1\text{H}$  NMR spectrum of 6-Hydroxy-2-methoxyxanthone (**3**)



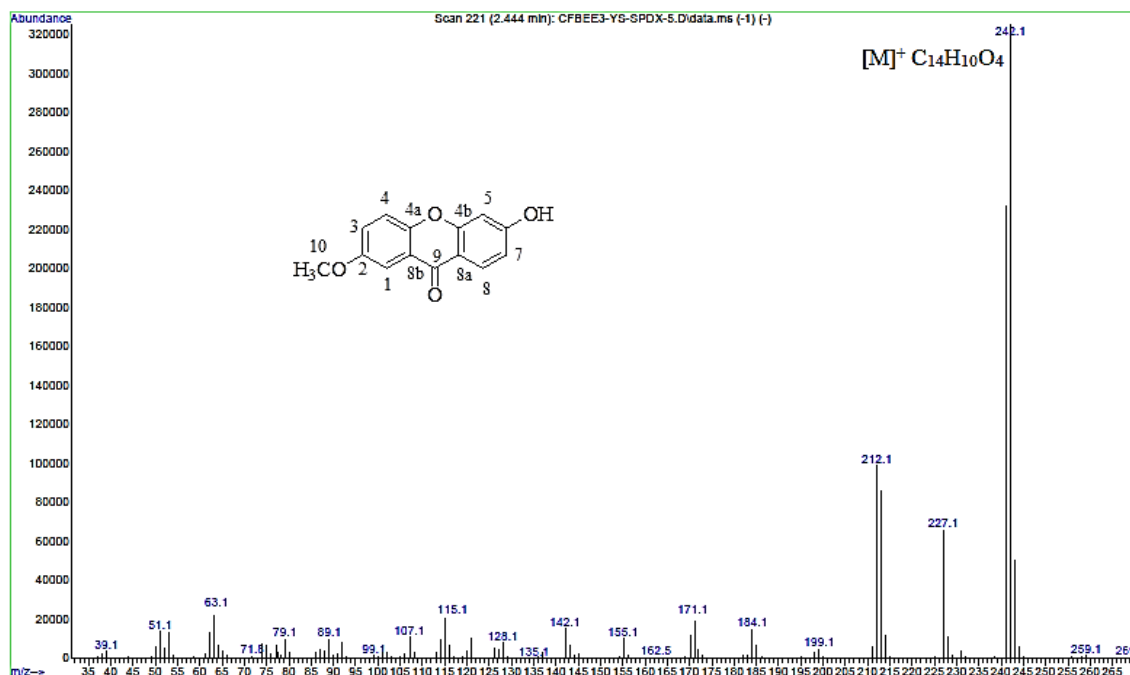
**S15:**  $^1\text{H}$  NMR spectrum of 6-Hydroxy-2-methoxyxanthone (**3**) (Expansion)



S16: DEPTQ spectrum of 6-Hydroxy-2-methoxyxanthone (3)

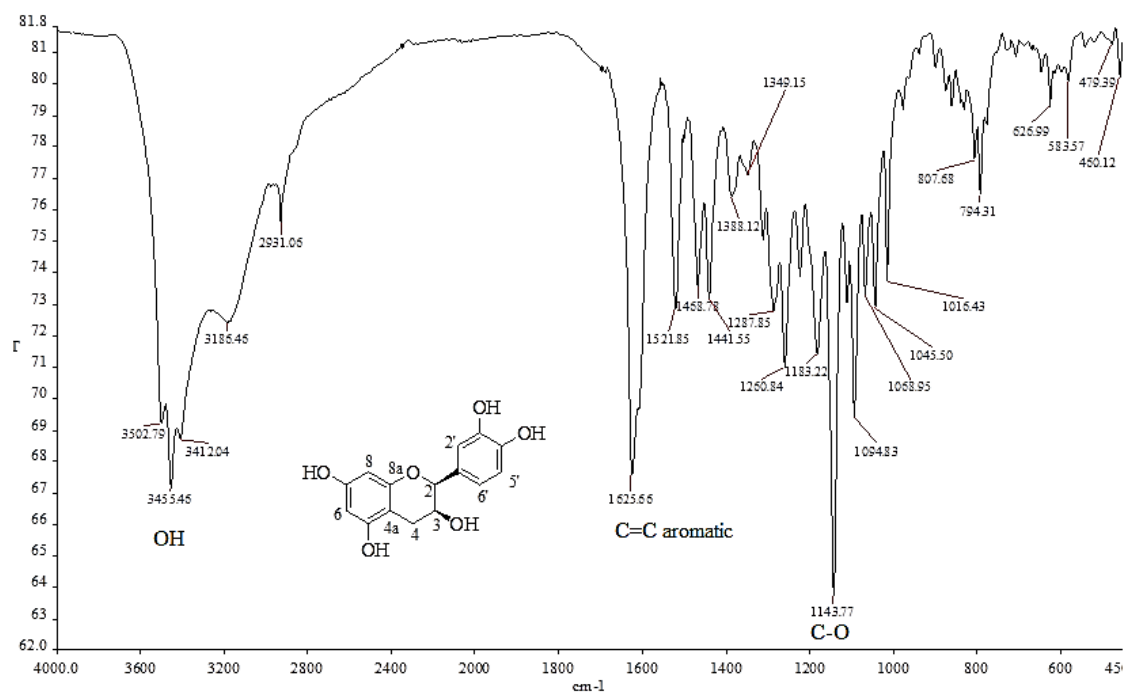


S17: DEPTQ\_Q spectrum of 6-Hydroxy-2-methoxyxanthone (3)

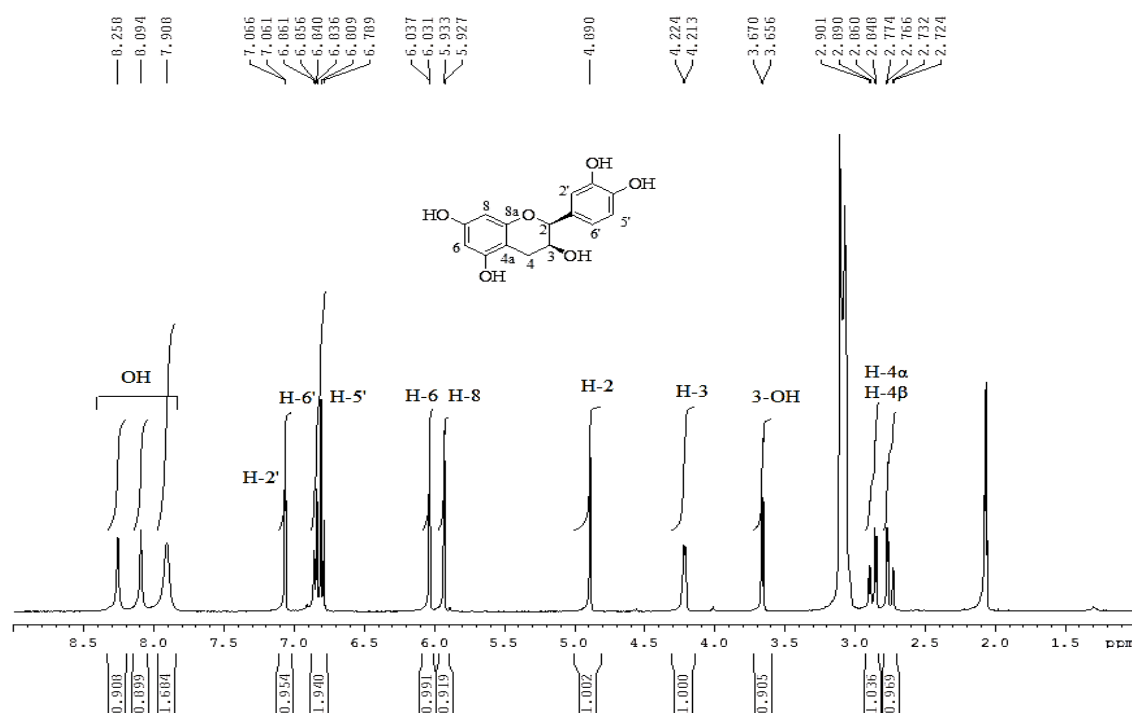


**S18:** EIMS spectrum of 6-Hydroxy-2-methoxyxanthone (**3**)

*ent*-Epicatechin (**4**): Pale brown amorphous;  $R_f$  0.25 (*n*-Hex:EtOAc, 1:1); m.p 235 – 236°C;  $[\alpha]_D^{25} +48.9^\circ$  (*c* 0.067, MeOH); IR (KBr pellet)  $\nu_{\max}$   $\text{cm}^{-1}$ : 3455 (OH), 2931 ( $sp^3$  CH), 1625 (C=C aromatic), 1143 and 1016 (C-O);  $^1\text{H}$  NMR (400 MHz,  $\text{CD}_3\text{COCD}_3$ ):  $\delta$  2.75 (1H, dd,  $J = 16.8$  and  $3.2$  Hz, H-4 $\alpha$ ), 2.86 (1H, dd,  $J = 16.8$  and  $4.4$  Hz, H-4 $\beta$ ), 3.66 (1H, d,  $J = 5.6$  Hz, 3-OH), 4.22 (1H, br s, H-3), 4.89 (1H, s, H-2), 5.93 (1H, d,  $J = 2.4$  Hz, H-8), 6.03 (1H, d,  $J = 2.4$  Hz, H-6), 6.79 (1H, d,  $J = 8.0$  Hz, H-5'), 6.85 (1H, dd,  $J = 8.0$  and  $2.0$  Hz, H-6'), 7.06 (1H, d,  $J = 2.0$  Hz, H-2'), 7.91 (2H, br s, 3'-OH and 4'-OH), 8.09 (1H, br s, 7-OH) and 8.26 (1H, br s, 5-OH);  $^{13}\text{C}$  NMR (100 MHz,  $\text{CD}_3\text{COCD}_3$ ):  $\delta$  28.2 (C-4 $\alpha$  and C-4 $\beta$ ), 66.0 (C-3), 78.5 (C-2), 94.7 (C-8), 95.3 (C-6), 98.9 (C-4 $\alpha$ ), 114.4 (C-2'), 114.6 (C-5'), 118.4 (C-6'), 131.3 (C-1'), 144.4 (C-4'), 144.5 (C-3'), 156.2 (C-8 $\alpha$ ), 156.6 (C-5) and 156.7 (C-7); EIMS (% rel int):  $m/z$  291 (4)  $[M+H]^+$ ,  $m/z$  290 (22),  $[M]^+$  ( $C_{15}H_{14}O_6$ ), 152 (40), 139 (100), 123 (46).

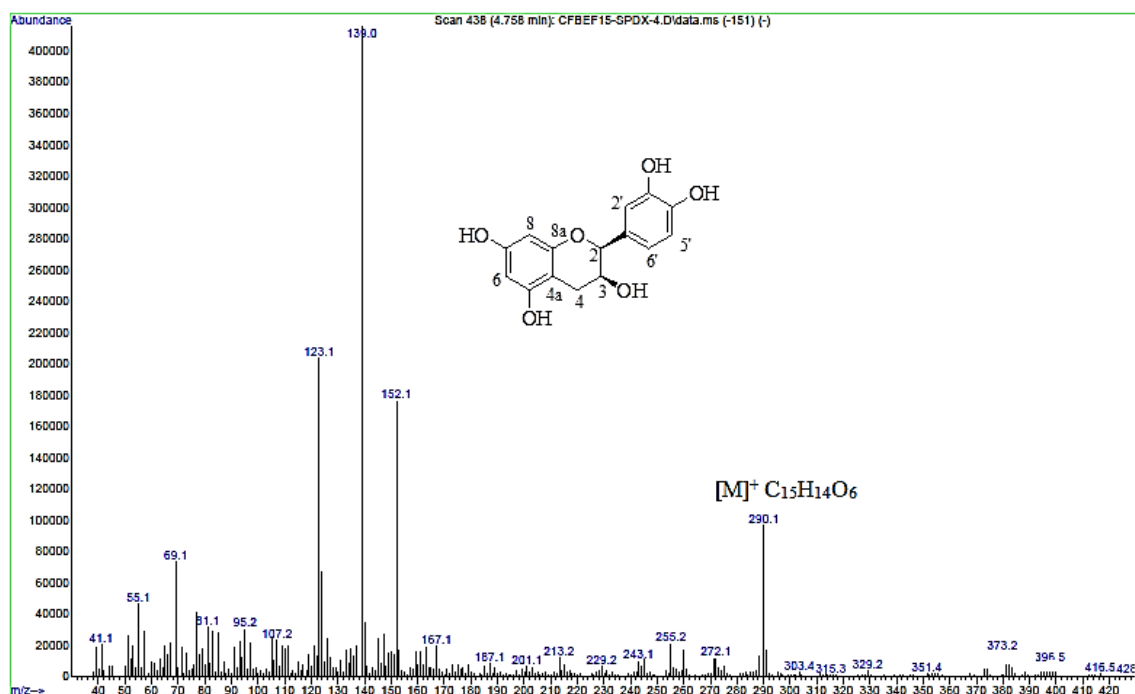
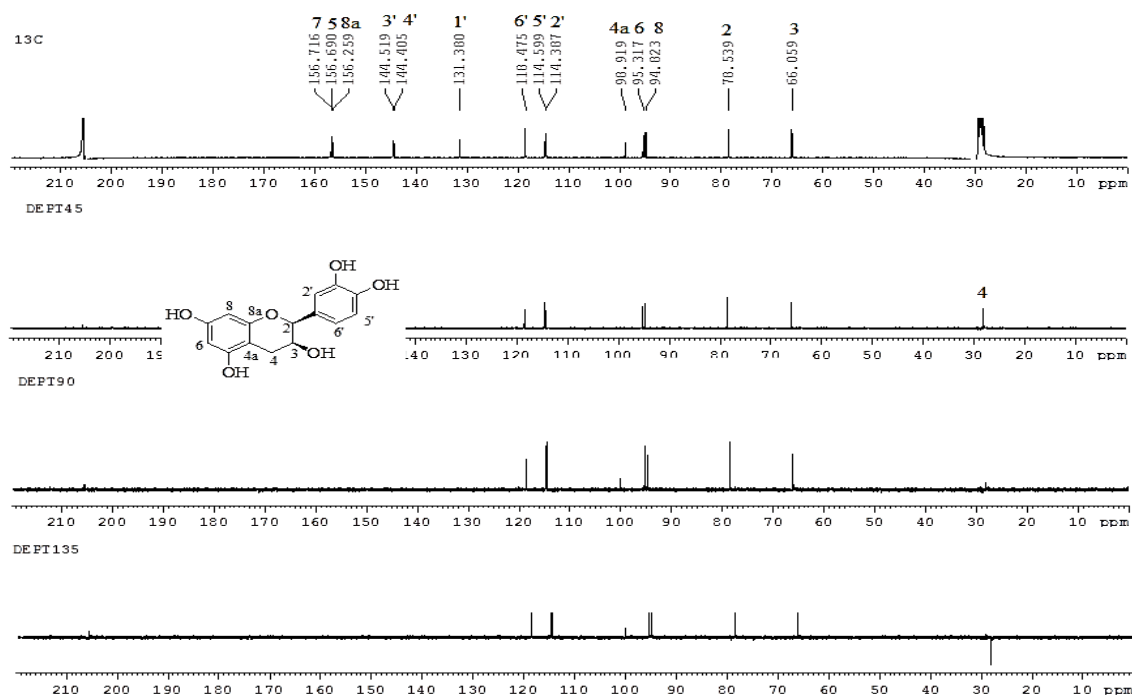


**S19:** IR spectrum of *ent*-Epicatechin (4)

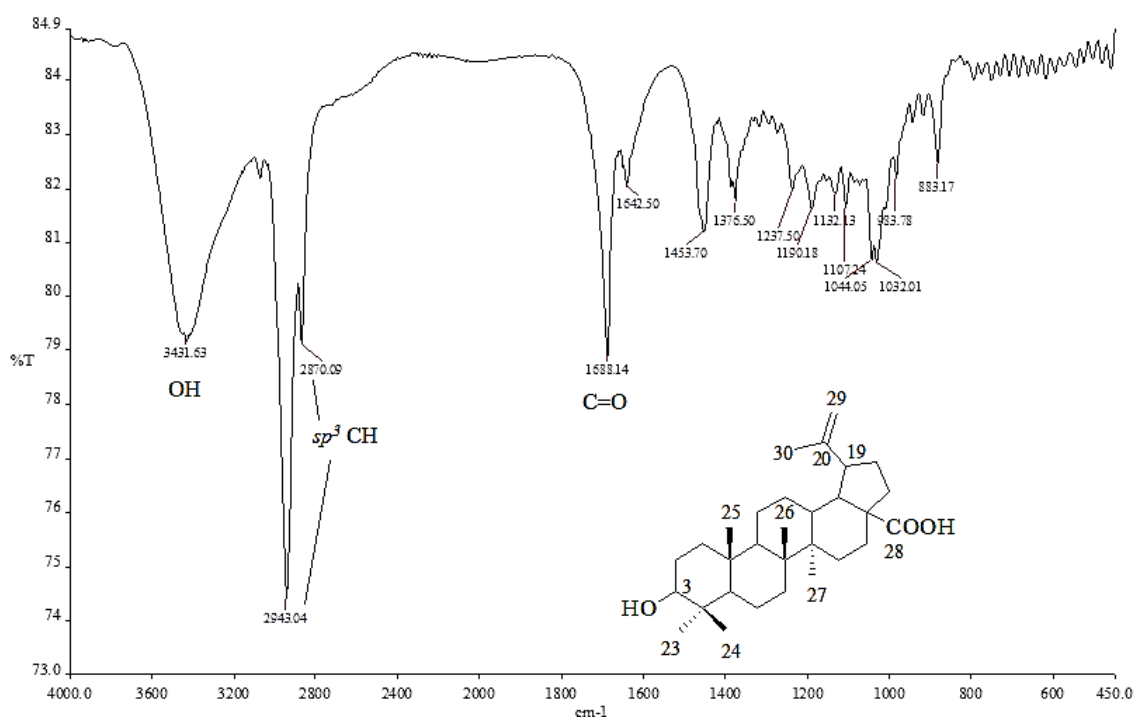


**S20:**  $^1\text{H}$  NMR spectrum of *ent*-Epicatechin (4)

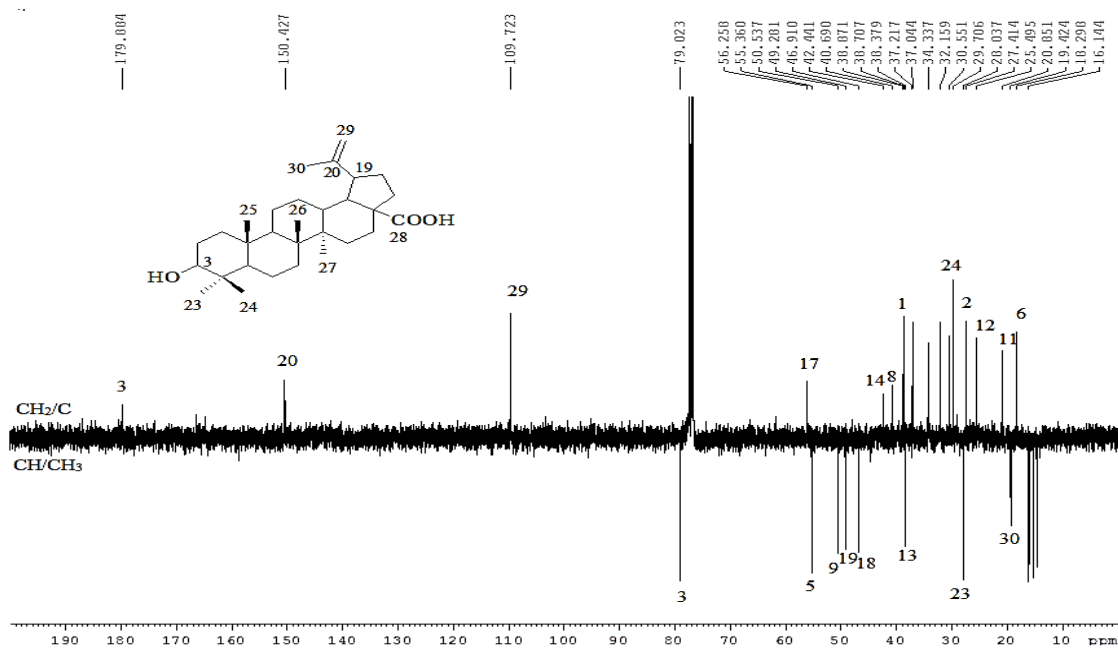
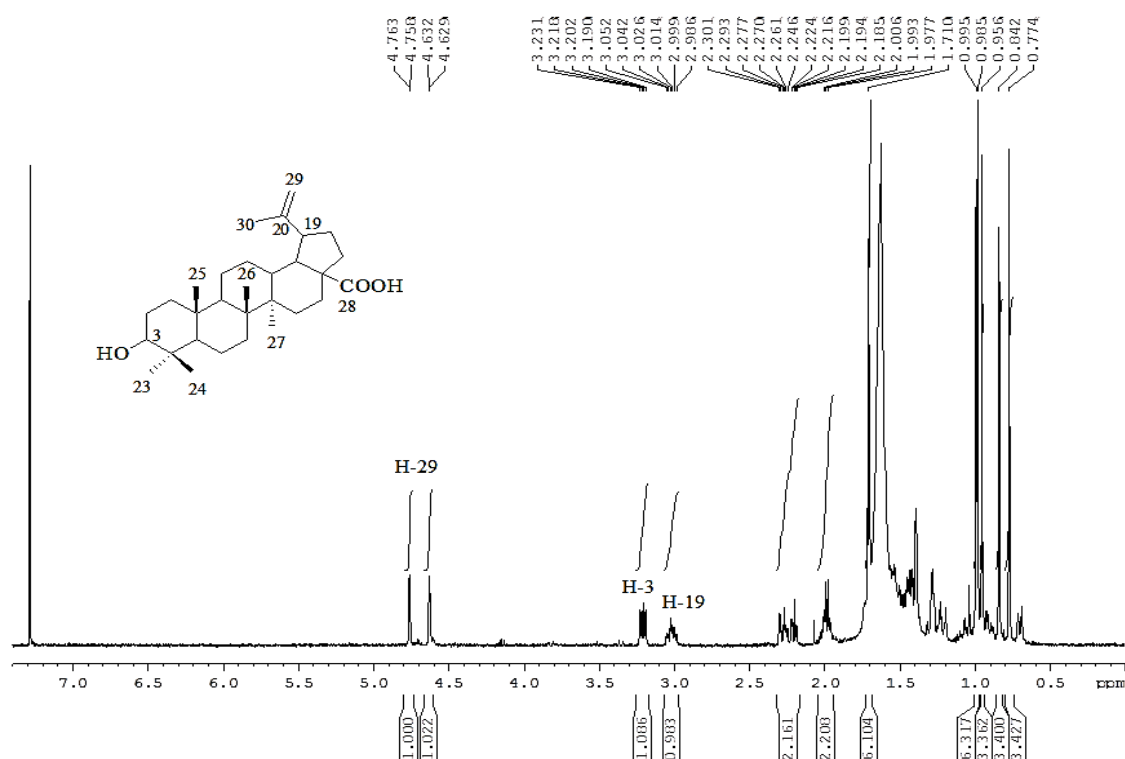


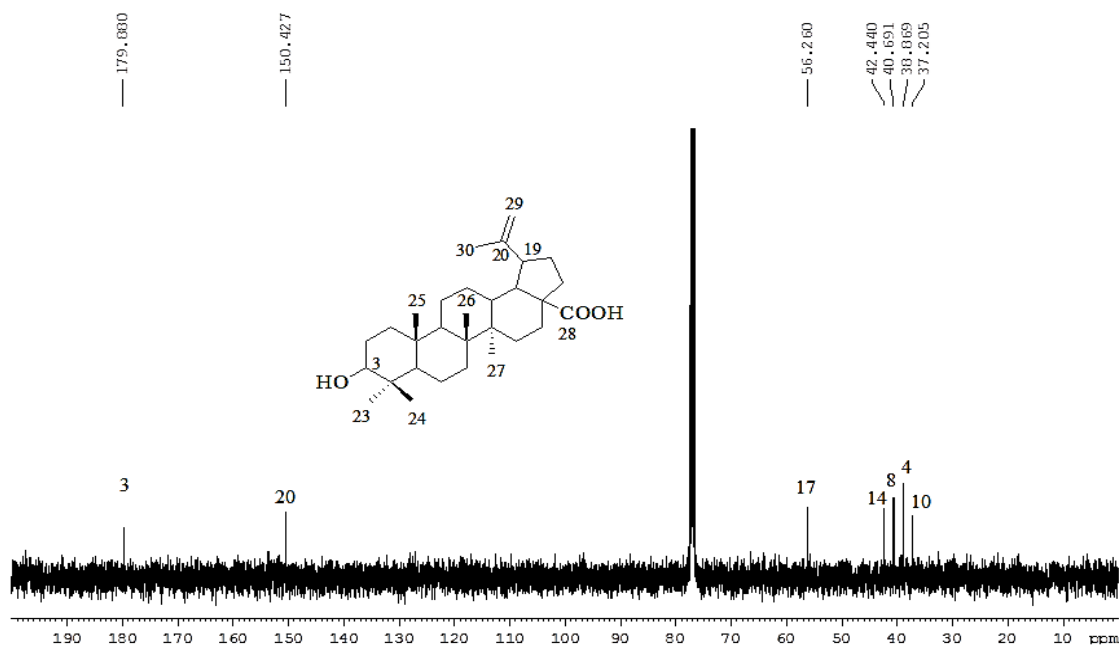


Betulinic acid (**5**): White solid;  $R_f$  0.55 ( $n$ -Hex: EtOAc, 3:2); m.p 311 – 313°C; IR (KBr pellet)  $\nu_{\max}$   $\text{cm}^{-1}$ : 3431 (OH), 2943 and 2870 ( $sp^3$  CH) and 1688 (C=O carboxylic acid);  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ ):  $\delta$  0.77 (3H, s, H-24), 0.84 (3H, s, H-25), 0.96 (3H, s, H-27), 0.98 (3H, s, H-23), 0.99 (3H, s, H-26), 1.71 (3H, s, H-30), 3.00 (1H, m, H-19), 3.21 (1H, dd,  $J = 4.8$  and 11.2 Hz, H-3), 4.76 (1H, br s,  $\text{H}_a$ -29), 4.73 (1H, br s,  $\text{H}_b$ -29);  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ ):  $\delta$  14.7 (C-27), 15.4 (C-24), 16.0 (C-25), 16.1 (C-26), 18.3 (C-6), 19.4 (C-30), 20.9 (C-11), 25.5 (C-12), 27.4 (C-2), 28.0 (C-23), 29.7 (C-21), 30.5 (C-15), 32.2 (C-16), 34.3 (C-7), 37.0 (C-22), 37.2 (C-10), 38.4 (C-13), 38.7 (C-1), 38.9 (C-4), 40.7 (C-8), 42.4 (C-14), 46.9 (C-18), 49.2 (C-19), 50.5 (C-9), 55.4 (C-5), 56.3 (C-17), 79.0 (C-3), 109.7 (C-29), 150.4 (C-20), 179.9 (C-28); EIMS (% rel int):  $m/z$  456 (7)  $[\text{M}]^+$  ( $\text{C}_{30}\text{H}_{48}\text{O}_3$ ), 248 (24), 203 (45), 189 (100).

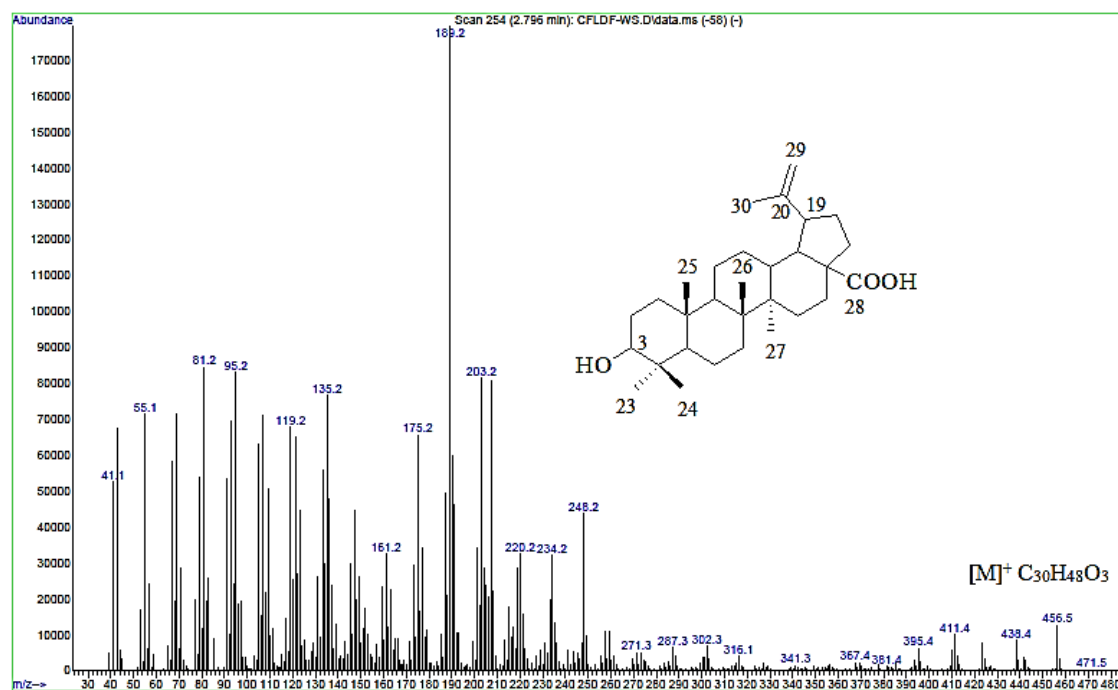


**S23:** IR spectrum of Betulinic acid (**5**)



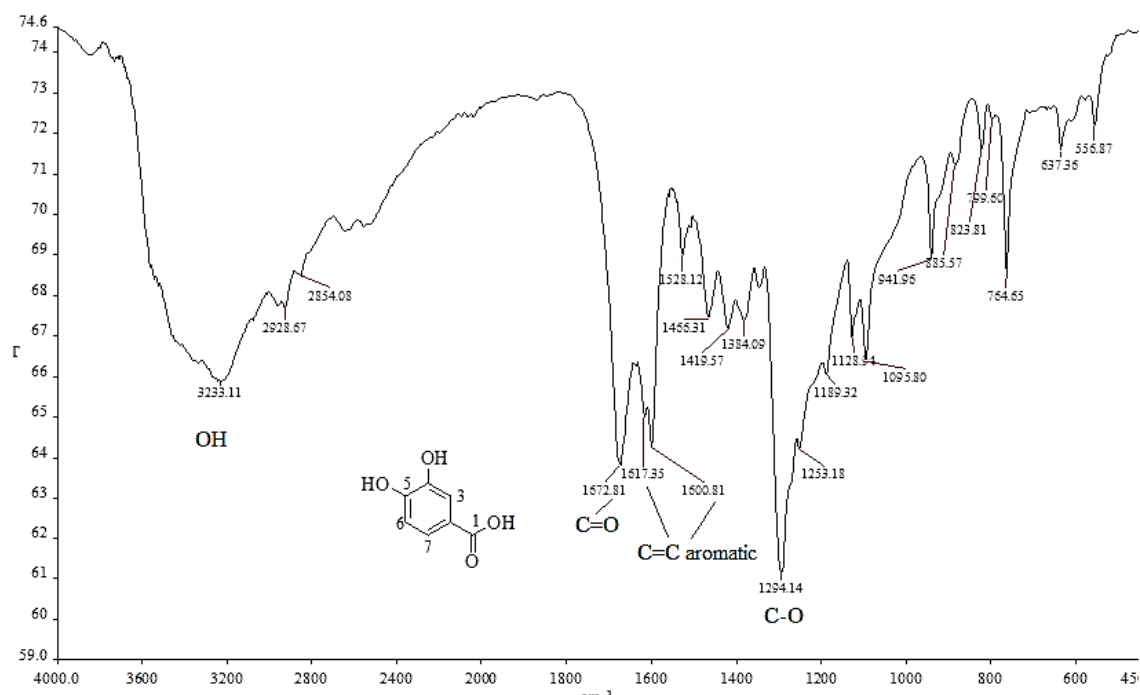


S26: DEPTQ-Q spectrum of Betulinic acid (5)

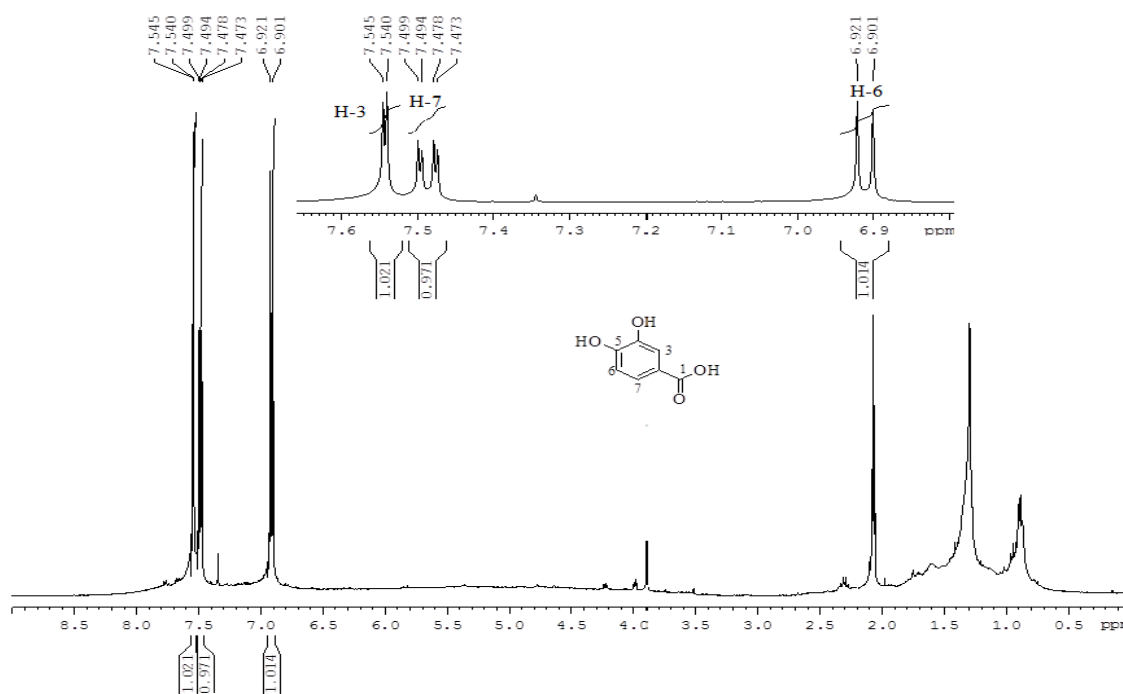


S27: EIMS spectrum of Betulinic acid (5)

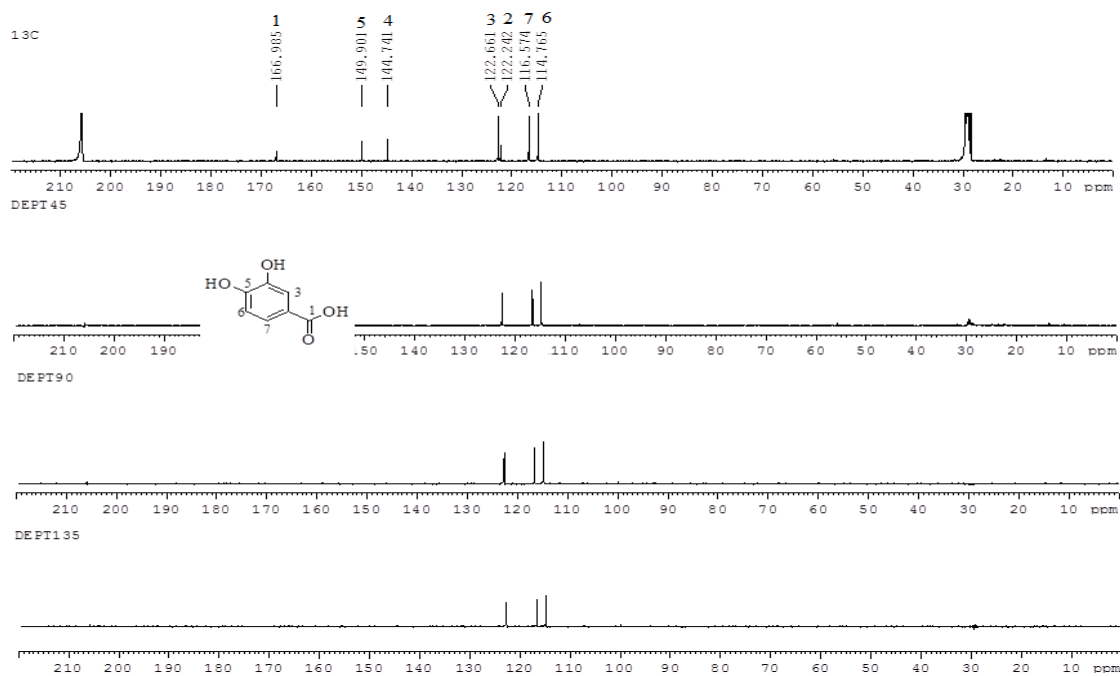
Protocatechuic acid (**6**): Yellow needle;  $R_f$  0.33 ( $n$ -Hex:EtOAc, 1:4); m.p 197 – 198°C; IR (KBr pellet)  $\nu_{\max}$   $\text{cm}^{-1}$ : 3244 (OH), 1673 (conjugated C=O carboxylic acid), 1617 and 1600 (C=C aromatic) and 1294 and 1095 (C-O);  $^1\text{H}$  NMR (400 MHz,  $\text{CD}_3\text{COCD}_3$ ):  $\delta$  6.91 (1H, d,  $J$  = 8.0 Hz, H-6), 7.49 (1H, dd,  $J$  = 8.0 and 2.0 Hz, H-7), 7.54 (1H, d,  $J$  = 2.0 Hz, H-3);  $^{13}\text{C}$  NMR (100 MHz,  $\text{CD}_3\text{COCD}_3$ ):  $\delta$  114.8 (C-6), 116.6 (C-7), 122.2 (C-2), 122.7 (C-3), 144.7 (C-4), 149.9 (C-5) and 166.9 (C-1); EIMS (% rel int):  $m/z$  154 (84)  $[\text{M}]^+$  ( $\text{C}_7\text{H}_6\text{O}_4$ ), 137 (100), 109 (24).



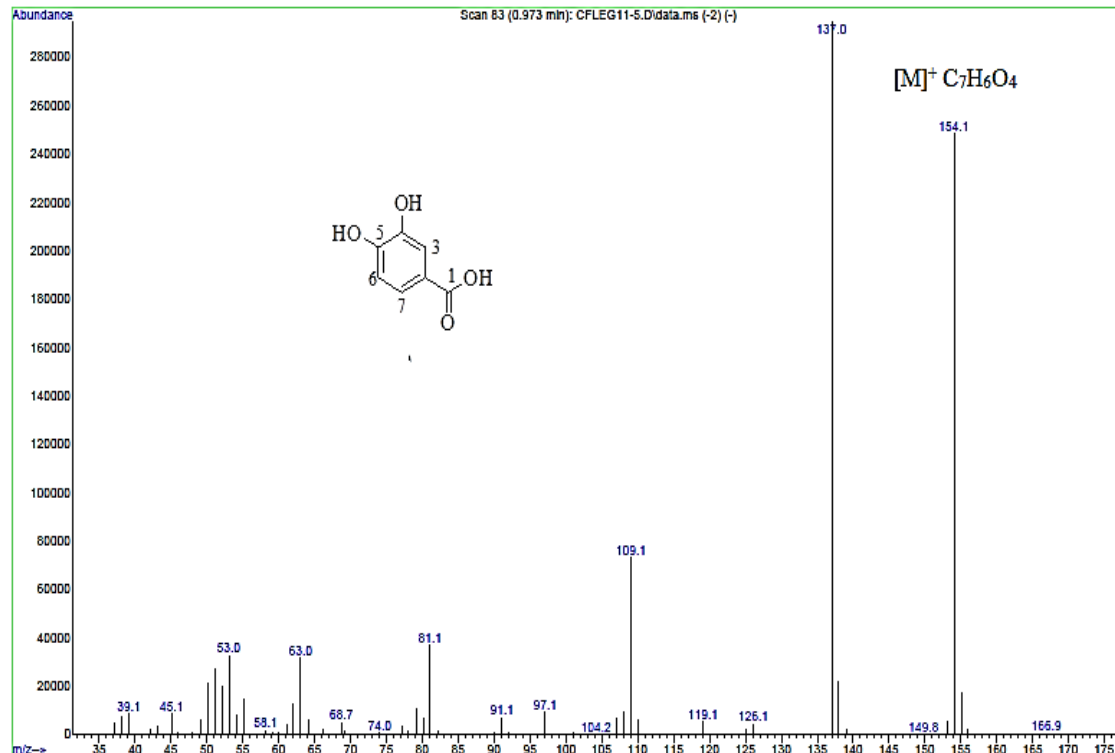
**S28:** IR spectrum of Protocatechuic acid (**6**)



**S29:** <sup>1</sup>H NMR of Protocatechuic acid (6)

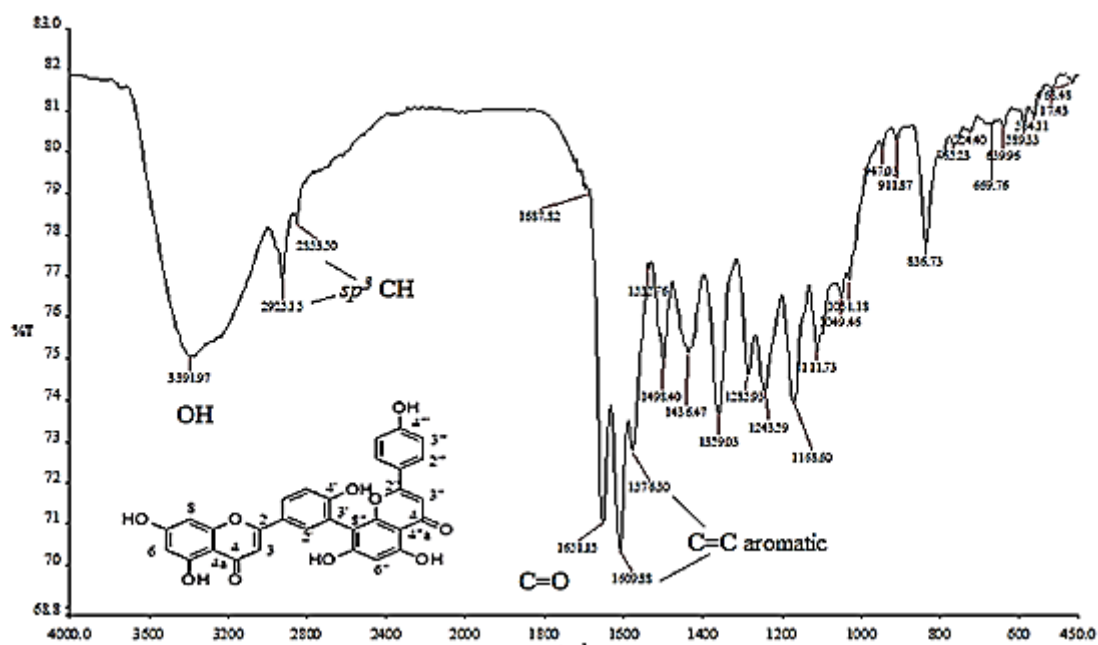


**S30:** <sup>13</sup>C/DEPT spectra of Protocatechuic acid (6)

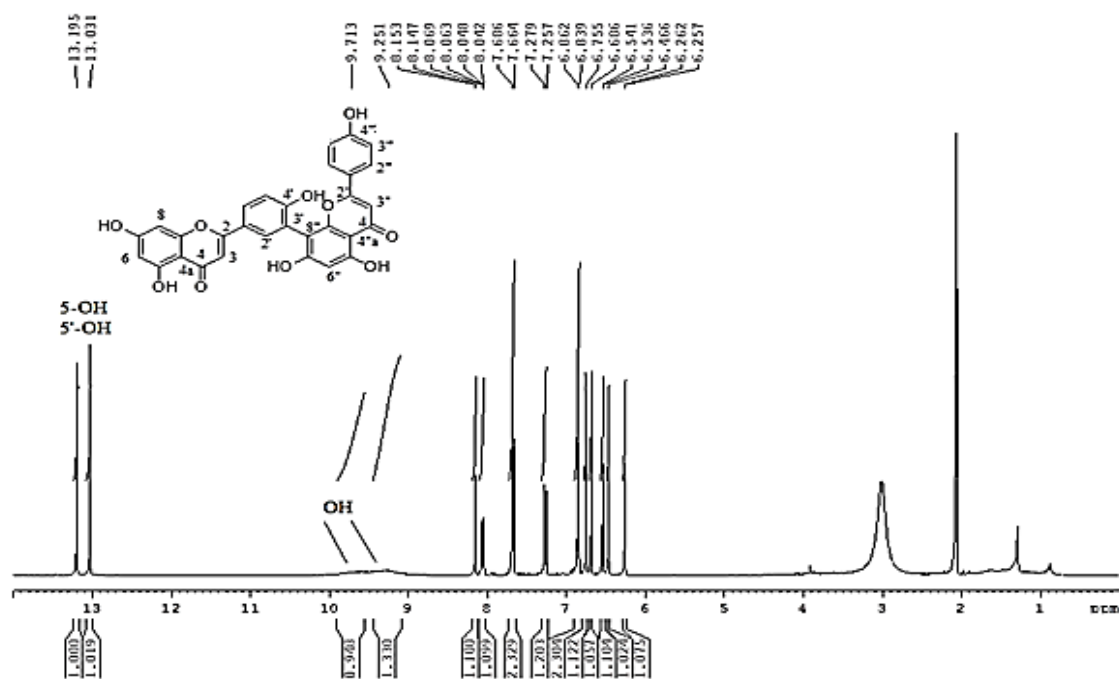


**S31:** EIMS spectrum of Protocatechuic acid (**6**)

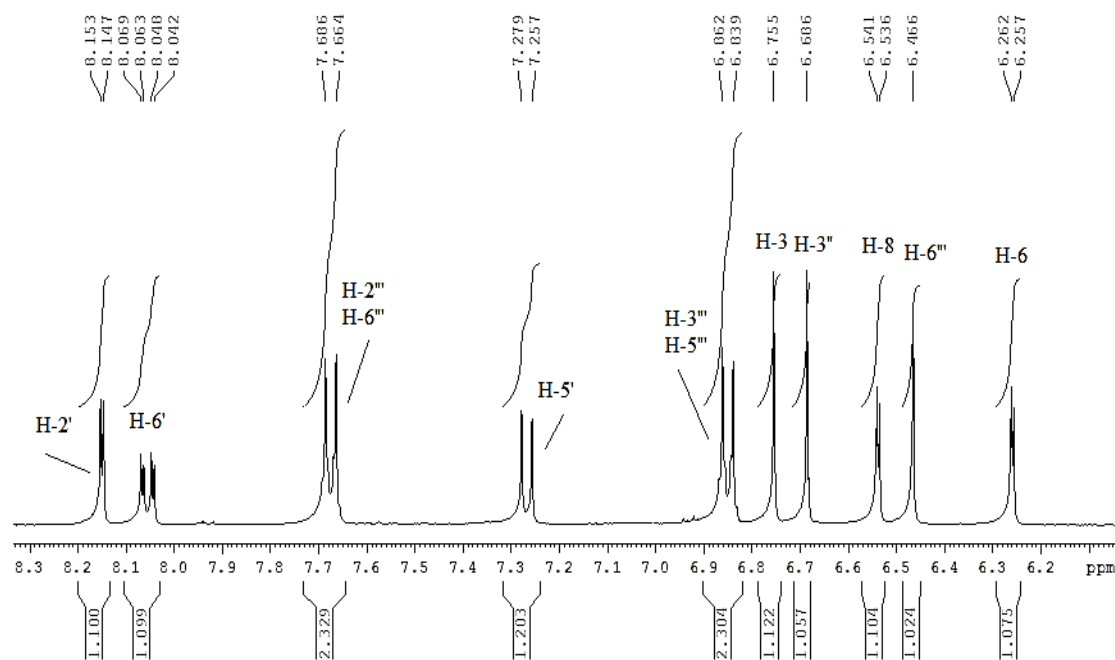
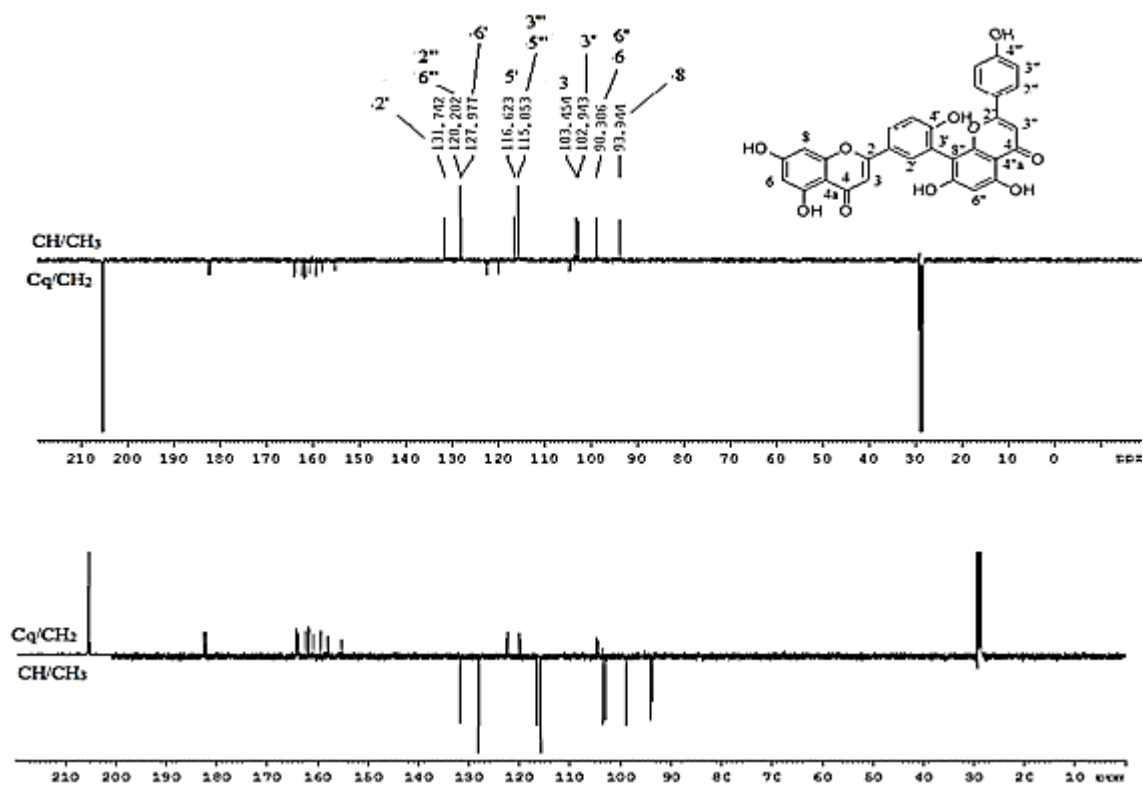
Amentoflavone (**7**): Yellow amorphous;  $R_f$  0.50 (*n*-Hex:EtOAc, 1:4); m.p 254 – 255°C; IR (KBr pellet)  $\nu_{\max}$   $cm^{-1}$ : 3392 (OH), 1651 (chelate C=O ketone), 1609 and 1576 (C=C aromatic) and 1168 and 1111 (C-O);  $^1H$  NMR (400 MHz,  $CD_3COCD_3$ ):  $\delta$  6.26 (1H, d,  $J = 2.4$  Hz, H-6), 6.47 (1H, s, H-6''), 6.54 (1H, d,  $J = 2.0$  Hz, H-8), 6.70 (1H, s, H-3''), 6.76 (1H, s, H-3), 6.86 (2H, d,  $J = 8.8$  Hz, H-3''' and H-5'''), 7.27 (1H, d,  $J = 8.8$  Hz, H-5'), 7.69 (2H, d,  $J = 8.8$  Hz, H-2''' and H-6'''), 8.07 (1H, dd,  $J = 8.8$  and 2.4 Hz, H-6'), 8.16 (1H, d,  $J = 2.4$  Hz, H-2'), 9.26 (1H, br s, 4'-OH), 9.73 (1H, br s, 4'''-OH), 13.05 (1H, s, 5-OH) and 13.21 (1H, s, 5''-OH).  $^{13}C$  NMR (100 MHz,  $CD_3COCD_3$ ):  $\delta$  93.9 (C-8), 98.9 (C-6 and C-6''), 102.9 (C-3''), 103.4 (C-3), 103.5 (C-8''), 104.5 (C-4a), 104.6 (C-4''a), 115.9 (C-3''' and C-5'''), 116.6 (C-5'), 119.9 (C-3'), 122.4 (C-1'), 122.5 (C-1'''), 127.9 (C-6'), 128.3 (C-2''' and C-6'''), 131.7 (C-2'), 155.2 (C-8''a), 157.9 (C-8a), 159.4 (C-4'), 161.0 (C-5''), 161.7 (C-5), 161.9 (C-7''), 162.5 (C-4'''), 164.0 (C-2 and C-7), 164.2 (C-2''), 182.2 (C-4) and 182.6 (C-4''); ESIMS (% rel int):  $m/z$  537 (100)  $[M-H]^+$  ( $C_{30}H_{18}O_{10}$ ).



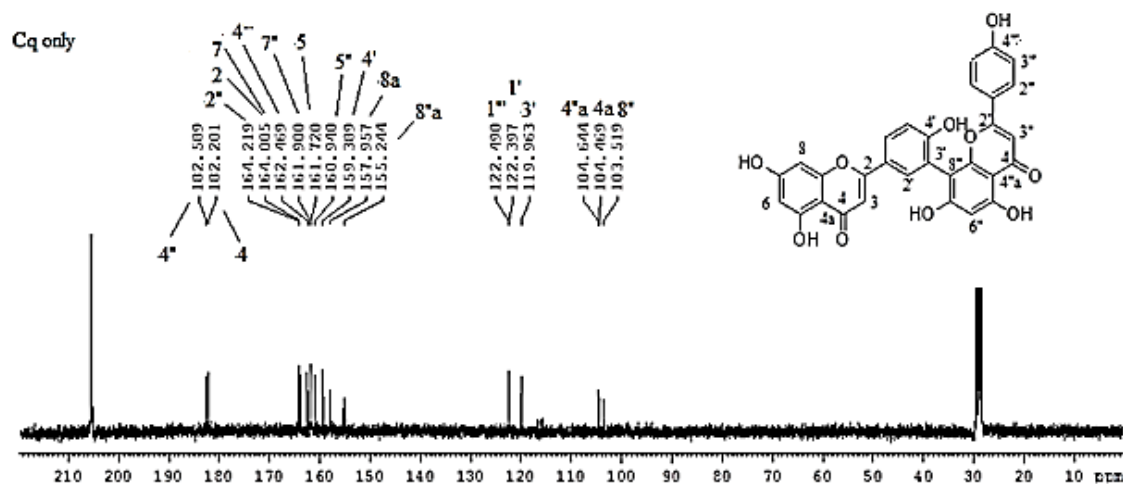
S32: IR spectrum of Amentoflavone (7)

S33:  $^1\text{H}$  NMR of Amentoflavone (7)

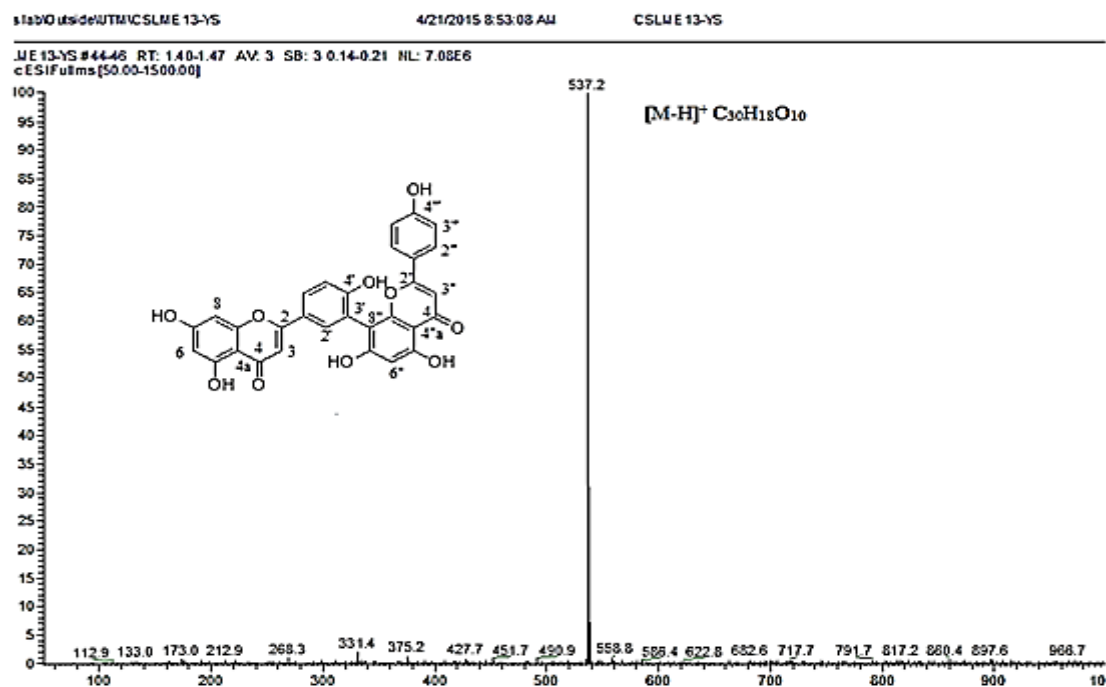


S34: <sup>1</sup>H NMR of Amentoflavone (7) (Expansion)

S35: DEPTQ spectra of Amentoflavone (7)



S36: DEPTQ-Q spectrum of Amentoflavone (7)



S37: ESIMS spectrum of Amentoflavone (7)

## References

- [1] A. A. Kaikabo and J. N. Eloff (2011). Antibacterial activity of two biflavonoids from *Garcinia livingstonei* leaves against *Mycobacterium smegmatis*, *J. Ethnopharmacol.* **138**, 253–255.
- [2] T. Mosmann (1983). Rapid colorimetric assay for cellular growth and survival: Application to proliferation and cytotoxicity assays, *J. Immunol. Methods*, **65**, 55–63.
- [3] M. Taher, D. Susanti, M. F. Rezali, F. S. A. Zohri, S. J.A. Ichwan, S.I. Alkhamaiseh and F. Ahmad (2012). Apoptosis, antimicrobial and antioxidant activities of phytochemicals from *Garcinia malaccensis* Hk.f., *Asian Pac. J. Trop. Med.* **5**, 136–141.
- [4] A. S. Ahmed, E. E. Elgorashi, N. Moodley, L. J. McGaw, V. Naidoo and J. Eloff (2012). The antimicrobial, antioxidative, anti-inflammatory activity and cytotoxicity of different fractions of four South African Bauhinia species used traditionally to treat diarrhoea, *J. Ethnopharmacol.* **143**, 826–839.
- [5] S. Perumal, S. Pillai, L. W. Cai, R. Mahmud and S. Ramanathan (2012). Determination of minimum inhibitory concentration of *Euphorbia hirta* (L.) extracts by tetrazolium microplate assay, *J. Nat. Prod.* **5**, 68–76.
- [6] C. Sarikurkcü, G. Zengin, M. Oskay, S. Uysal, R. Ceylan and A. Aktumsek (2015). Composition, antioxidant, antimicrobial and enzyme inhibition activities of two *Origanum vulgare* subspecies (subsp. *vulgare* and subsp. *hirtum*) essential oils, *Ind. Crops. Prod.* **70**, 178–184.
- [7] A. S. Sufian, K. Ramasamy, N. Ahmat, Z. A. Zakaria and M. I. M. Yusof (2013). Isolation and identification of antibacterial and cytotoxic compounds from the leaves of *Muntingia calabura* L., *J. Ethnopharmacol.* **146**, 198–204.